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Apport du séquençage dans les épidémies hospitalières

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Stephan Harbarth

62èmes journées Claude Bernard, 28 Nov 2019

Infection Control Programme, University of Geneva Hospitals and Faculty of Medicine, Geneva, Switzerland

Remerciements: Romain Martischang

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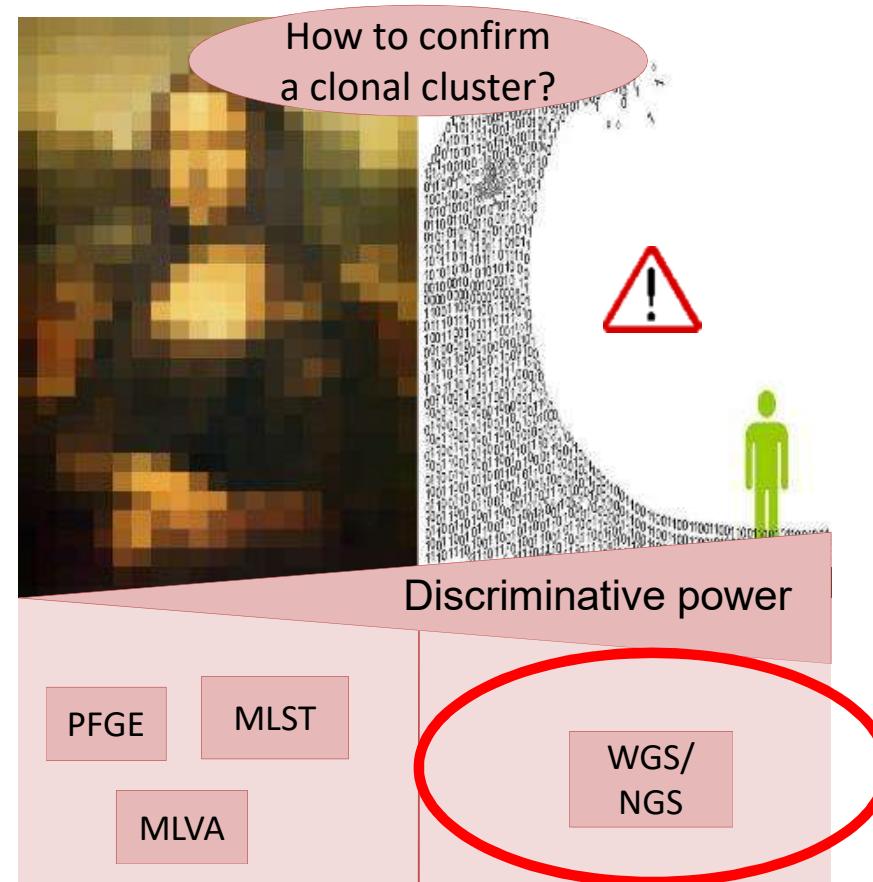
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Key Words:

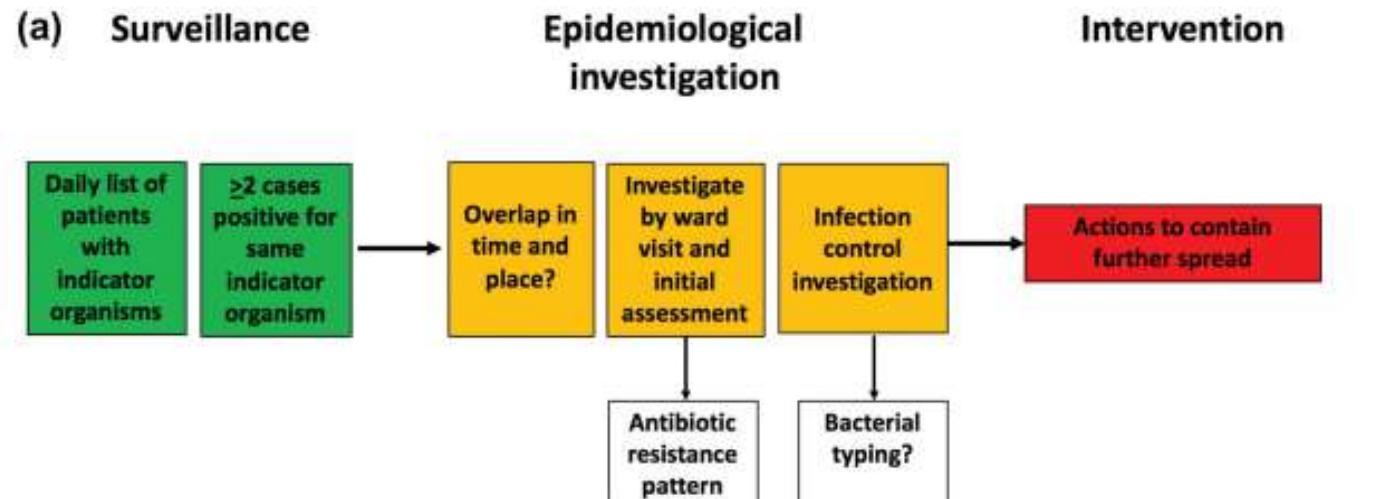
Hand hygiene;

World Health Organization;

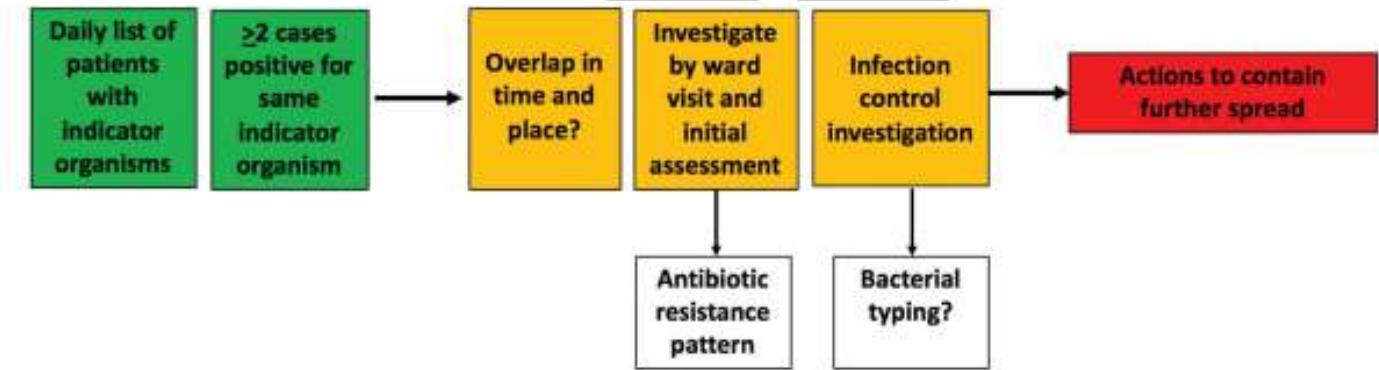
Detection and confirmation of a monoclonal nosocomial outbreak



Conventional approach to outbreak detection



NGS-based outbreak detection



(b)

Peacock et

Routine

Probability of outbreak

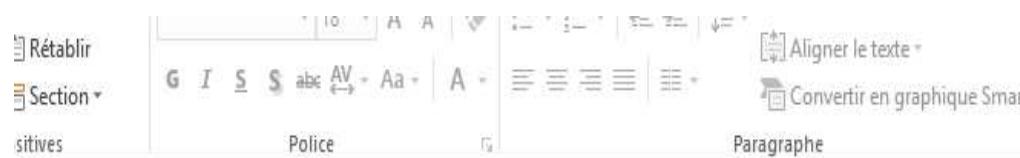
High

Actions to contain further spread

Making epidemiological inferences based on molecular data

We expect **epidemiologically linked isolates** to be **genetically identical** or similar, therefore:

We expect the bacterial population to have a clonal structure
=> Detection of **monoclonal clusters of isolates**.

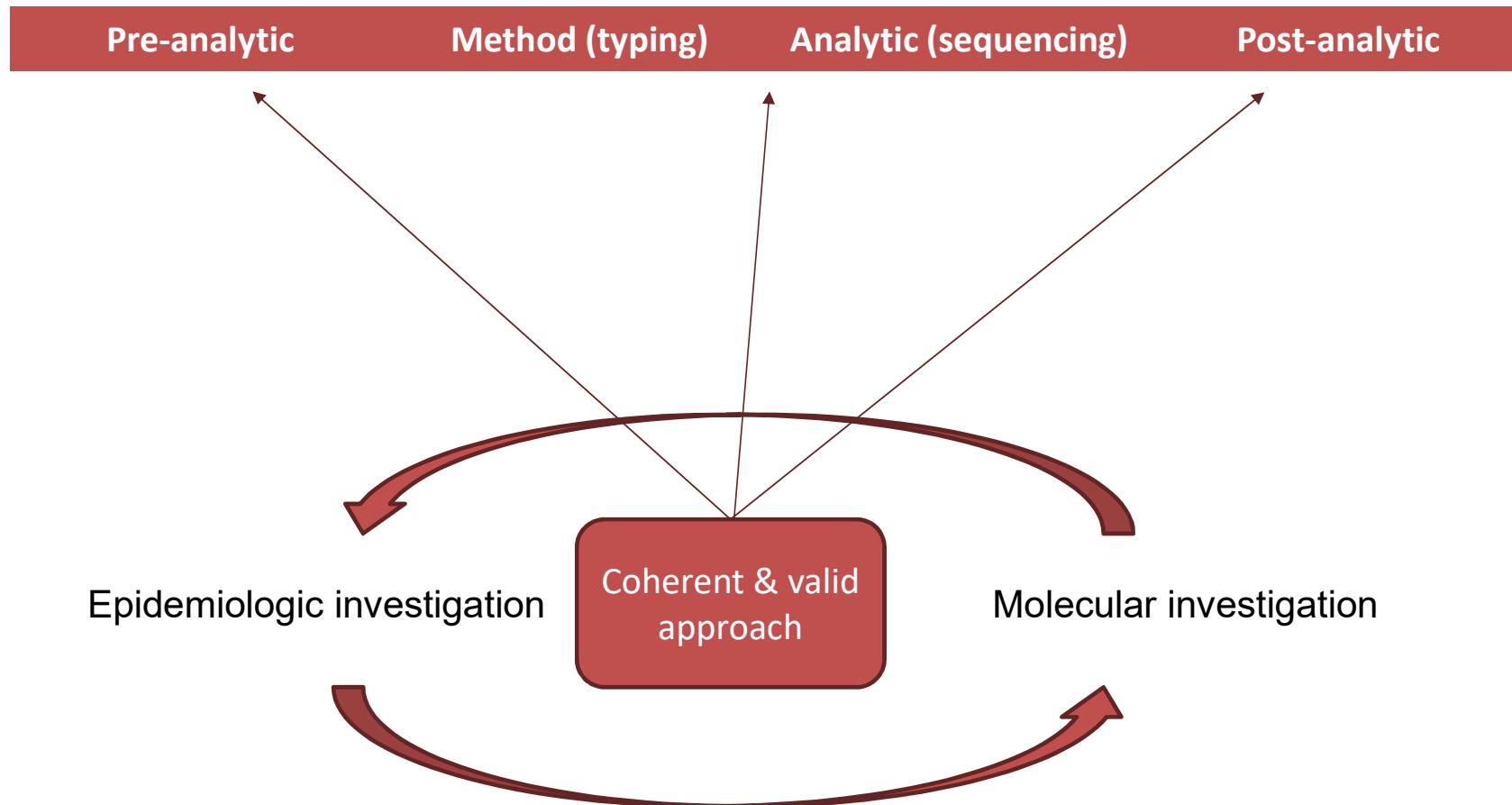


Has the time come to perform a paradigm shift towards routine NGS?



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L'ESSENTIEL, C'EST VOUS.



Indication for molecular typing?

- Do we have an epidemiologic hypothesis ?

→ Molecular fishing expedition?



Indication for molecular typing?

- Do I have an epidemiologic hypothesis ?

→ Molecular fishing expedition?

- Do I expect any impact on infection control interventions ? Or futile academic exercise ?

→ NGS: priority action item for changing preventive measures?



Genomic Surveillance Reveals Diversity of Multidrug-Resistant Organism Colonization and Infection: A Prospective Cohort Study in Liver Transplant Recipients

Nenad Macesic,^{1,2} Angela Gomez-Simmonds,¹ Sean B. Sullivan,^{1,3} Marla J. Giddins,^{1,3} Samantha A. Ferguson,¹ Gautam Korakavi,¹ David Leeds,¹ Sarah Park,¹ Kevin Shim,¹ Madeleine G. Sowash,¹ Melanie Hofbauer,¹ Ryan Finkel,¹ Yue Hu,¹ Jared West,¹ Nora C. Toussaint,^{4,a} William G. Greendyke,¹ Benjamin A. Miko,¹ Marcus R. Pereira,¹ Susan Whittier,⁵ Elizabeth C. Verna,⁶ and Anne-Catrin Uhlemann^{1,3}

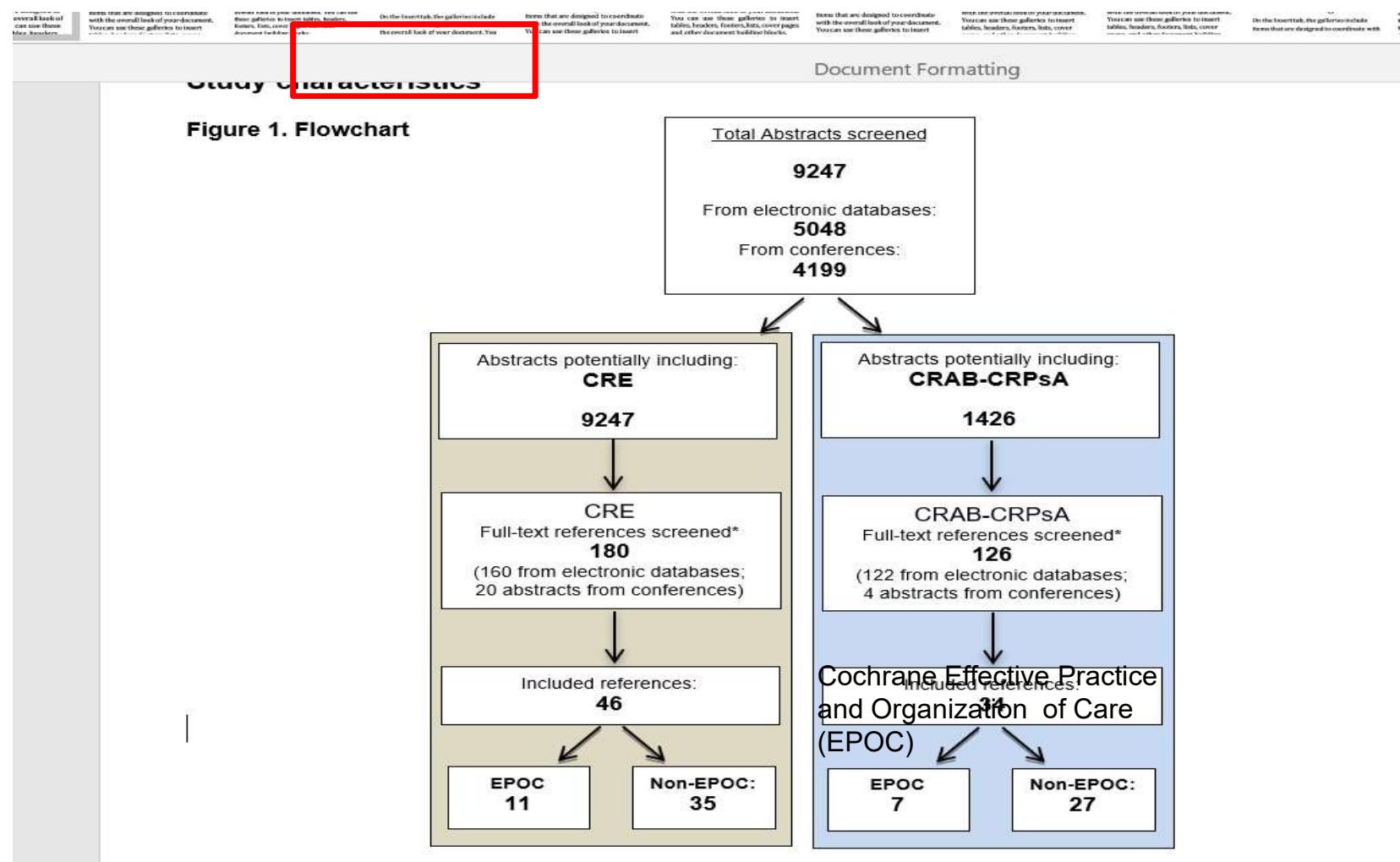
Results. We collected 998 stool samples and 119 rectal swabs from 128 patients. MDRO colonization was detected in 86 (67%) patients at least once and was significantly associated with subsequent MDRO infection (0 vs 19.8%, $P = .002$). Child-Turcotte-Pugh score at LT and duration of post-LT hospitalization were independent predictors of both MDRO colonization and infection. Temporal dynamics differed between MDROs with respect to onset of colonization, clearance, and infections. We detected an unexpected diversity of CRE colonizing isolates and previously unrecognized transmission that spanned Ceph-RE and CRE phenotypes, as well as a cluster of *mcr-1*-producing isolates.

Conclusions. Active surveillance and WGS showed that MDRO colonization is a highly dynamic and complex process after LT. Understanding that complexity is crucial for informing decisions regarding MDRO infection control, use of therapeutic decolonization, and empiric treatment regimens.

WHO: CPE control

- No RCT or controlled study
- All EPOC-studies are from CRE-endemic countries: Israel, USA, Italy and Brazil
- All describe multi-faceted interventions
- EPOC: 10 interrupted time series studies
- Non-EPOC (N=36)
 - 17 Non-controlled before-after studies
 - 14 Before-after case-counts
 - 3 Modeling studies
 - 2 Longitudinal studies

CRE control review -- Flowchart



Infection control measures in high-quality CPE control studies

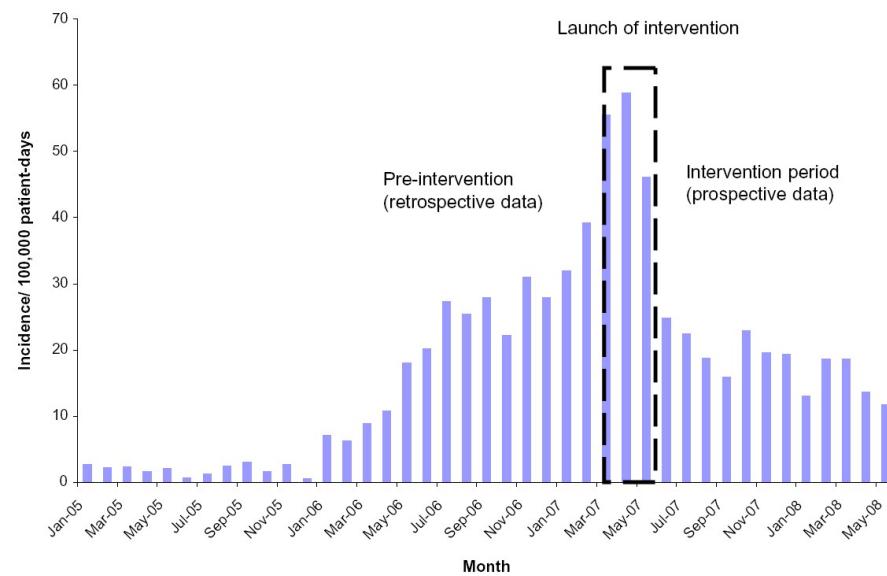
-- Systematic WHO review & meta-analysis --

Intervention	EPOC studies
Active surveillance	10/11
Contact precautions	10/11
Cohorting	9/11
Monitoring, audit and feedback	9/11
Patient isolation	9/11
Hand hygiene education & monitoring	6/11
Education	4/11
Antibiotic stewardship	4/11
Enhanced environmental cleaning	3/11
Daily chlorhexidine gluconate baths	3/11
Flagging positive patients in medical record (alerts)	3/11
Environmental surveillance	1/11
Temporary ward closure	1/11

Containment of a Country-wide Outbreak of Carbapenem-Resistant *Klebsiella pneumoniae* in Israeli Hospitals via a Nationally Implemented Intervention

Clinical Infectious Diseases 2011;52(7):1–8

Mitchell J. Schwaber,¹ Boaz Lev,² Avi Israeli,² Ester Solter,¹ Gill Smolan,¹ Bina Rubinovitch,¹ Itamar Shalit,¹ Yehuda Carmeli,¹ and the Israel Carbapenem-Resistant Enterobacteriaceae Working Group^a



MID 2018
Hall A
viewers

o Calendar

in overview

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gramme

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valuate

Stalking a lethal superbug by whole-genome sequencing and phylogenetics: Influence on unraveling a major hospital outbreak of carbapenem-resistant *Klebsiella pneumoniae*

Thorsten Kaiser MD ^a, Knut Finstermeier PhD ^a, Madlen Häntzsch MSc ^a, Sarah Faucheuix MD ^b, Martin Kaase MD ^c, Tim Eckmanns MD ^d, Sven Bercker MD, PhD ^e, Udo X. Kaisers MD, PhD ^e, Norman Lippmann MD ^{f,g}, Arne C. Rodloff MD, PhD ^{f,g}, Joachim Thiery MD, PhD ^a, Christoph Lübbert MD, PhD, DTM&H ^{g,h,*}

- **34-month outbreak in Leipzig University Hospital**
- **84/105 KPC-2-kp (ST258) available**
- **Index case transferred from Rhodes, Greece**
- **19 wards affected**

WGS & Spread of KPC-2

- Positioning pillow to maintain patient prone in ICU, might have been the link
- Additional cases after screening
- 34 median number of single nucleotide variants

Epidemiology

Pathways explained for 11
(12.4%) patients

WGS

Pathways explained for 15
extra patients & confirmed
5 of epidemiology

- Exact mode of transmission unknown for 63 (71%) patients

Indication for molecular typing?

- Do I have an epidemiologic hypothesis ?
→ Molecular fishing expedition?
- Do I expect any impact on infection control interventions ? Or simply academic exercise ?
→ Priority action item for modified preventive measures?

Sampling?

- Do I have a strong, robust sampling strategy? (Who, How, When)
- Avoid:
- detection bias
 - selection bias
 - misclassification bias
- Adequate screening for asymptomatic carriers
 - If possible, select the right colonies by selective cultures (multiresistant organisms).
 - How many morphologically similar isolates to sample from the same clinical culture ?

Multiple Variants of *Klebsiella pneumoniae* Producing Carbapenemase in One Patient

- 82 yr man
- Admitted to the same hospital 21 times (2011-2015)
- Often grouped with patients colonized with KPC-producing Enterobacteriaceae (KPE)
- Numerous antibiotic courses

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Jewish General Hospital
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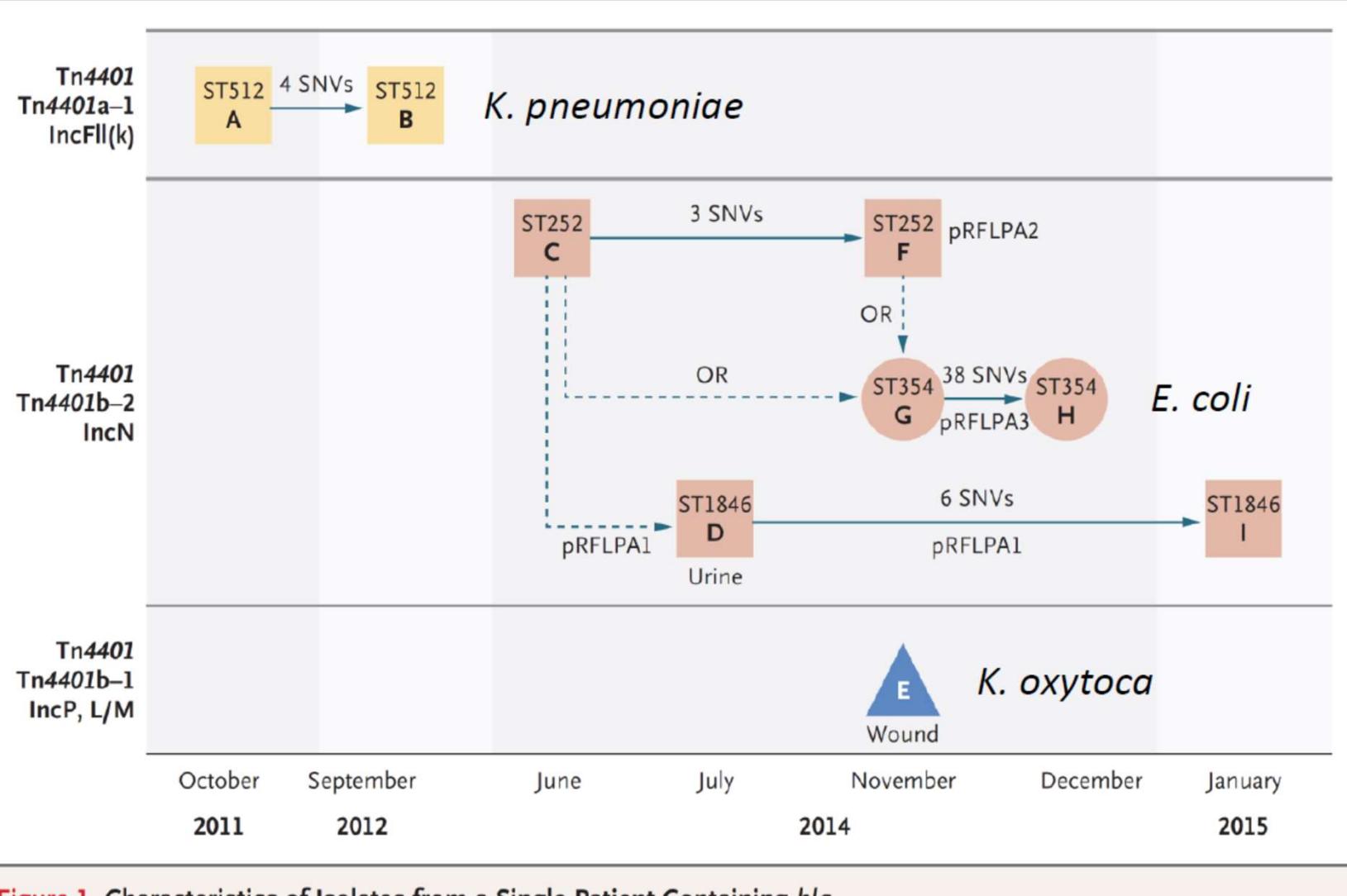
Mulvey et al. NEJM 2016; 375: 2408-10

Mulvey et al. NEJM 2016; 375: 2408-10



- 14 KPE isolates identified
- 9 available for sequencing:
 - 7 rectal swabs
 - 1 urine
 - 1 wound swab
- 3 species (*K. pneumoniae*, *K. oxytoca*, and *E. coli*) identified, including:
 - 3 different *K. pneumoniae* sequence types
 - 3 different incompatibility (Inc) group plasmids carrying Tn4401

Mulvey et al. NEJM 2016; 375: 2408-10



- Multiple isolates may have arisen from:
 - Spread of resistance element within the host and/or
 - Exposure to other KPE-colonized patients
- Limitations of outbreak investigations with traditional epidemiologic and molecular tools
- Genome sequencing necessary

Mulvey et al. NEJM 2016; 375: 2408-10

Pre-analytic

Method (typing)



Typing methods for outbreak investigations and epidemiologic surveillance

PFGE
MLVA



Various targets

POS: Established method
CONS: Little discriminatory power, cannot establish exact transmission routes

MLST



Selected loci

POS: Robust, reproducible method; allows to observe longterm trends
CONS: Little discriminatory power

Monocentric outbreak

Long-term
surveillance /
multicenter outbreaks

PFGE: électrophorèse en champs pulsé

MLVA: Analyse du polymorphisme de locus répétés

MLST: Analyse de locus multiples (séquençage ciblé)

RAPID COMMUNICATION

Outbreak of vancomycin-resistant *Enterococcus faecium* clone ST796, Switzerland, December 2017 to April 2018

Nasstasia Wassilew¹, Helena MB Seth-Smith^{2,3}, Eveline Rolli¹, Yvonne Fietze¹, Carlo Casanova⁴, Urs Führer⁵, Adrian Egli^{2,3}, Jonas Marschall¹, Niccolò Buetti¹

1. Department of Infectious Diseases, University Hospital Bern, Bern, Switzerland

2. Division of Clinical Microbiology, University Hospital Basel, Basel, Switzerland

3. Applied Microbiology Research, Department of Biomedicine, University of Basel, Basel, Switzerland

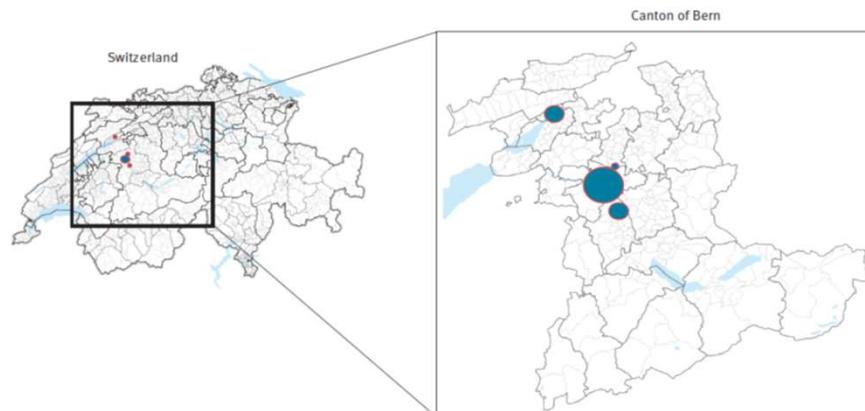
4. Institute for Infectious Diseases, University of Bern, Bern, Switzerland

5. Infectious Diseases Department, Biel Hospital, Biel, Switzerland

Correspondence: Niccolò Buetti (niccolo.buetti@gmail.com)

FIGURE 1

Distribution of vancomycin-resistant *Enterococcus faecium* ST796 in four different hospitals, Canton of Bern, Switzerland, 30 December 2017 to 30 April 2018 (n = 89)

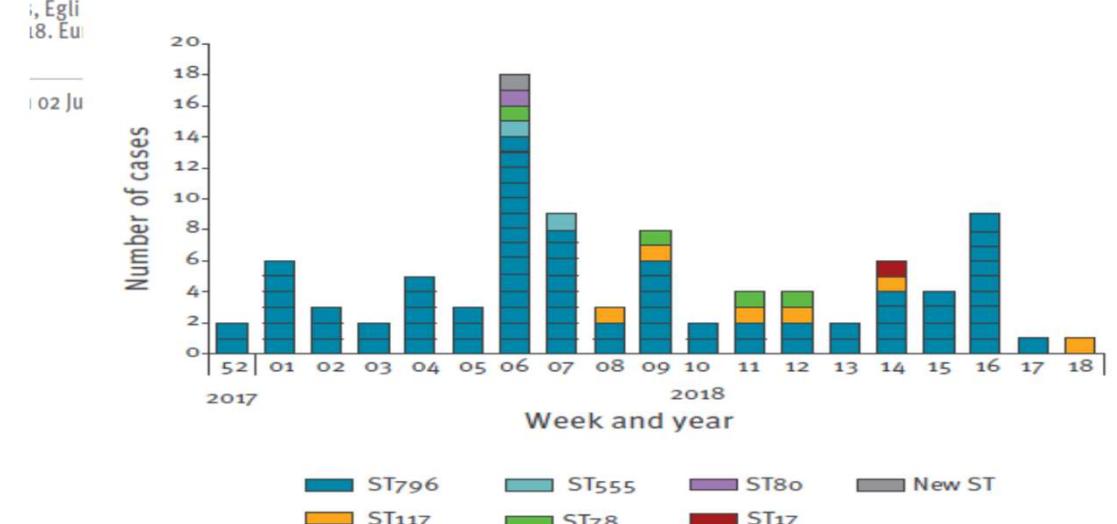


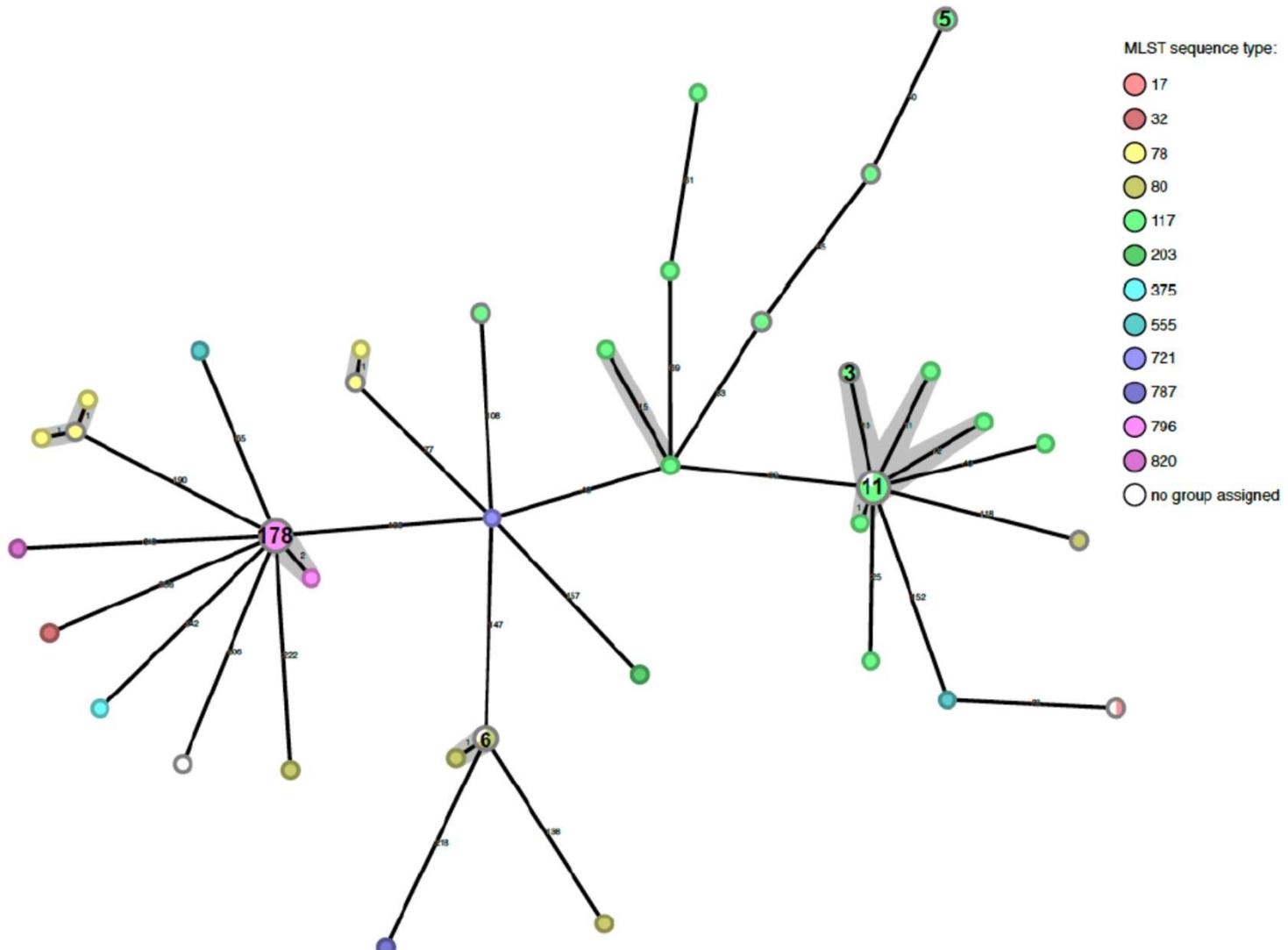
The largest outbreak at University hospital is shown by the largest circle.

EuroSurveillance 2018

FIGURE 3

Epidemic curve of vancomycin-resistant enterococci (VRE) cases by sequence type, Canton of Bern outbreak, Switzerland, December 2017–April 2018 (n = 89)





Pre-analytic

Method (typing)



Typing methods for outbreak investigations and epidemiologic surveillance



Various targets

POS: Established method
CONS: Little discriminatory power, cannot establish exact transmission routes

Monocentric outbreak



Selected loci

POS: Robust, reproducible method; allows to observe longterm trends
CONS: Little discriminatory power

Long-term Surveillance / Multicenter outbreaks



Core genome

POS: High discriminatory power
CONS: Still expensive and requires special analytical skills

Outbreaks or epidemiologic surveillance

WGS/NGS: séquençage de génome complet

VRE outbreak

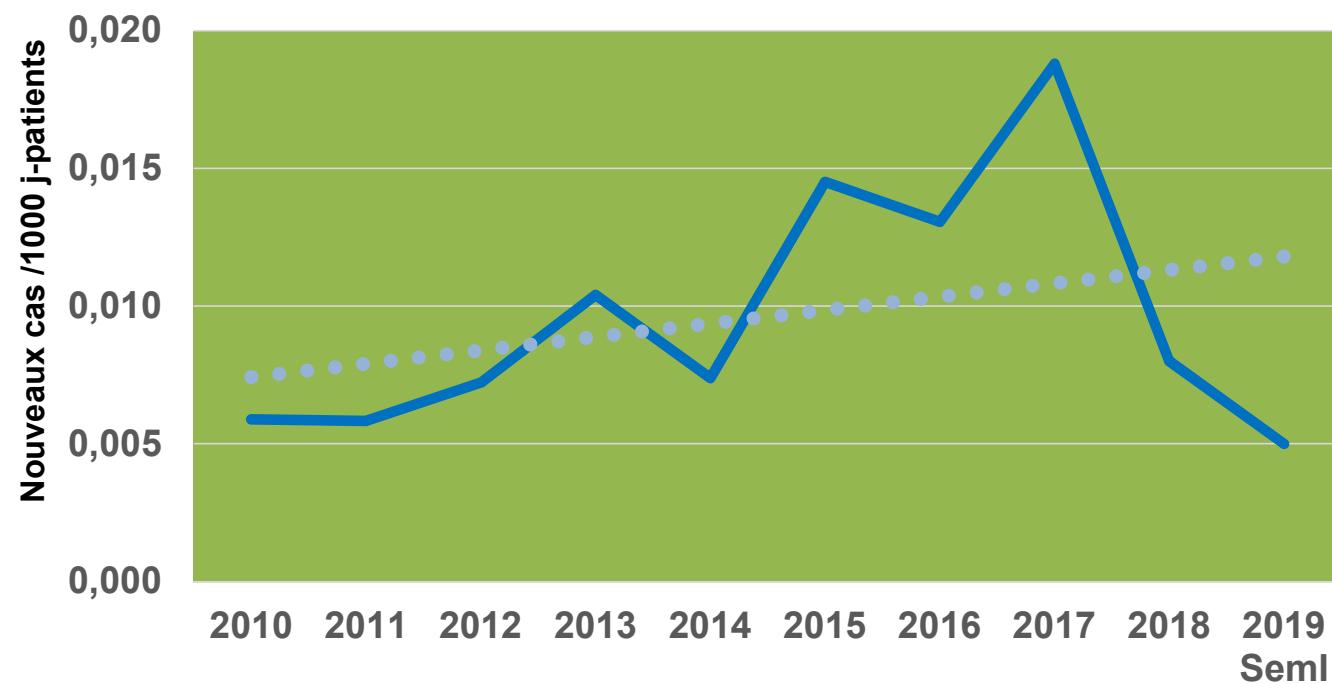
Geneva, surgical unit

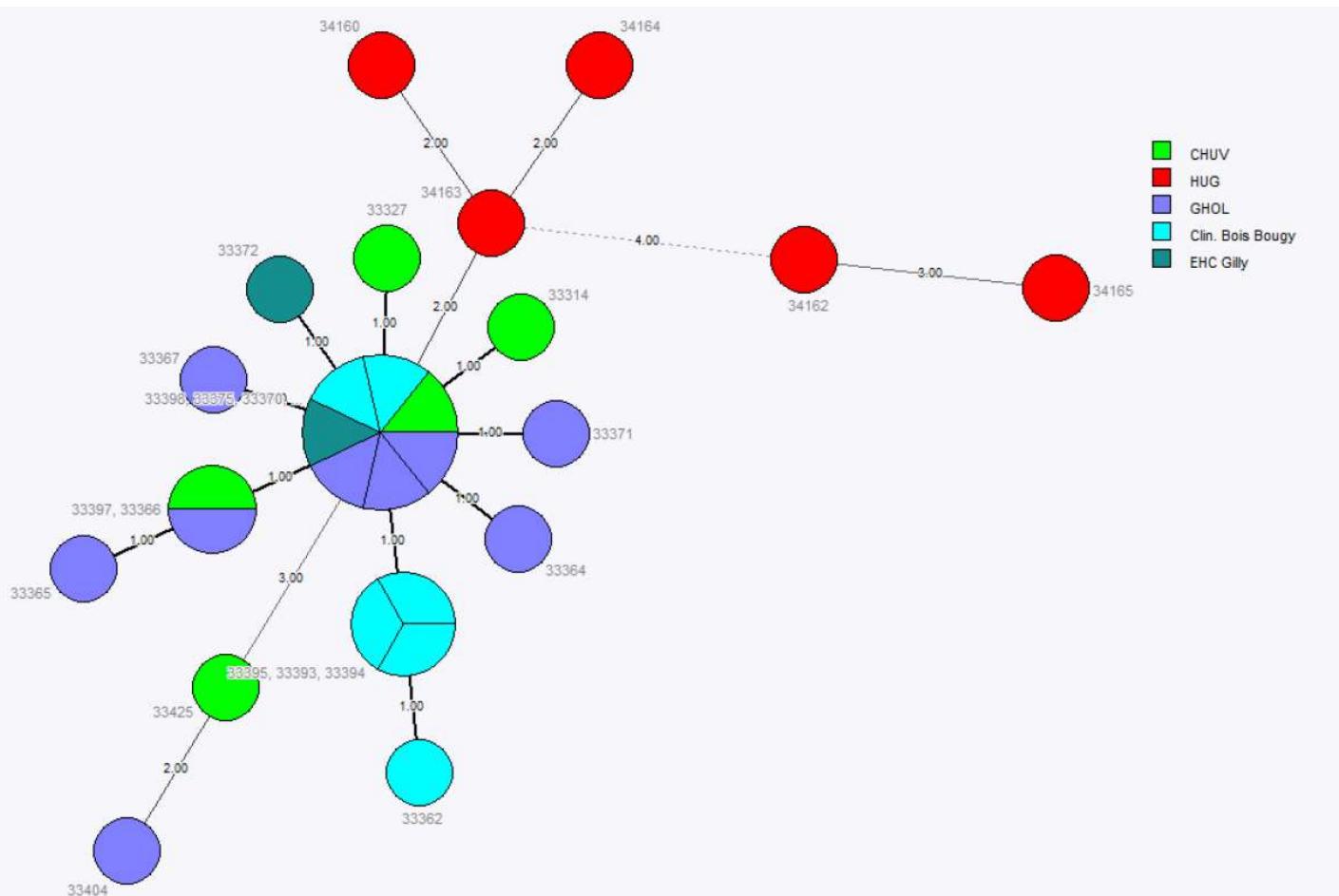


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Densité d'incidence de nouveaux cas VRE détectés après 48h d'admission HUG, Janvier 2010 – Juin 2019





Eclosions de VRE avec transmission régionale

Région du Lac Leman (2017)

Figure 1. Arbre de maximum de parcimonie entre les souches du cluster ST117-B. La distance entre chaque souche indique le nombre de loci de différence.

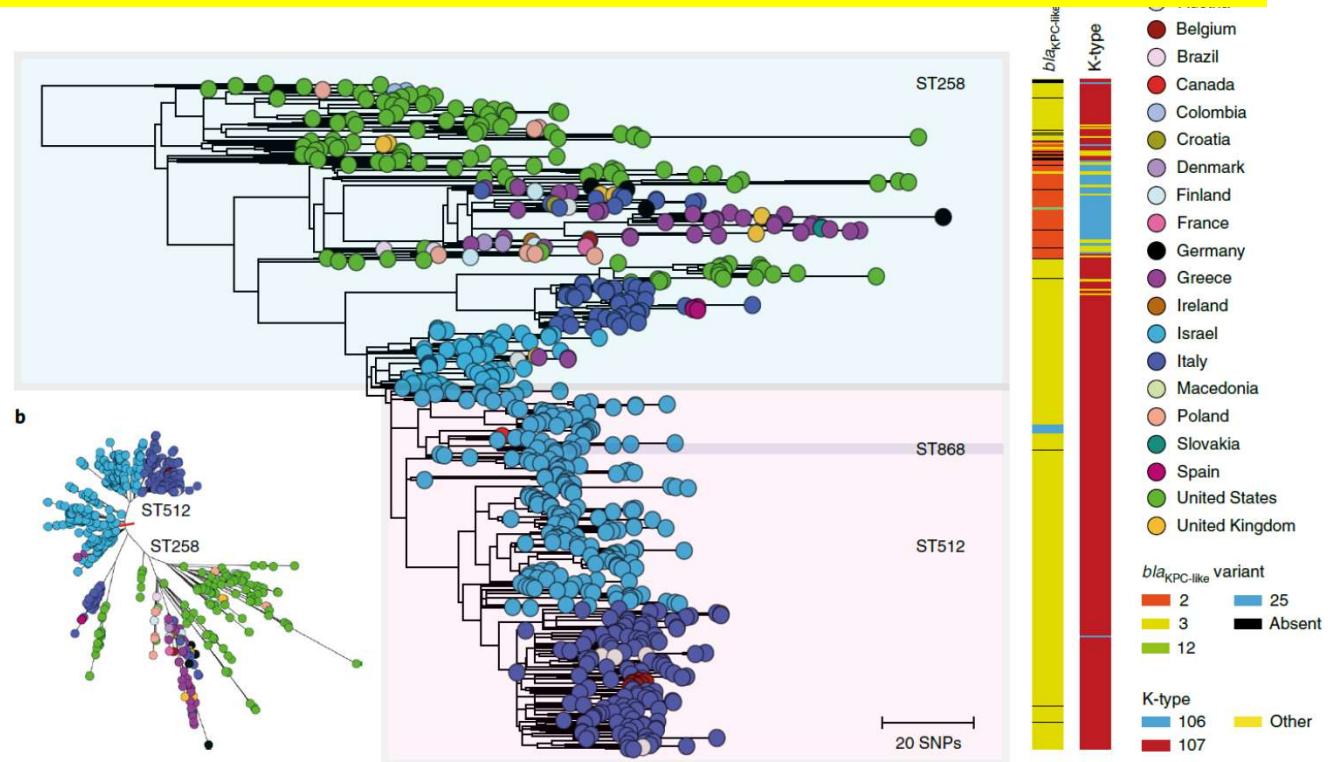
Courtesy: D. Blanc - CHUV

Global spread of three multidrug-resistant lineages of *Staphylococcus epidermidis*

Jean Y. H. Lee¹, Ian R. Monk¹, Anders Gonçalves da Silva^{2,3}, Torsten Seemann¹ ^{3,4}, Kyra Y. L. Chua⁵, Angela Kearns⁶, Robert Hill⁶, Neil Woodford⁶, Mette D. Bartels⁷, Birgit Strommenger⁸, Frederic Laurent⁹, Magali Dodémont¹⁰, Ariane Deplano¹⁰, Robin Patel¹¹, Anders R. Larsen¹², Tony M. Korman¹³, Timothy P. Stinear^{1,3,15} and Benjamin P. Howden¹ ^{2,3,14,15*}

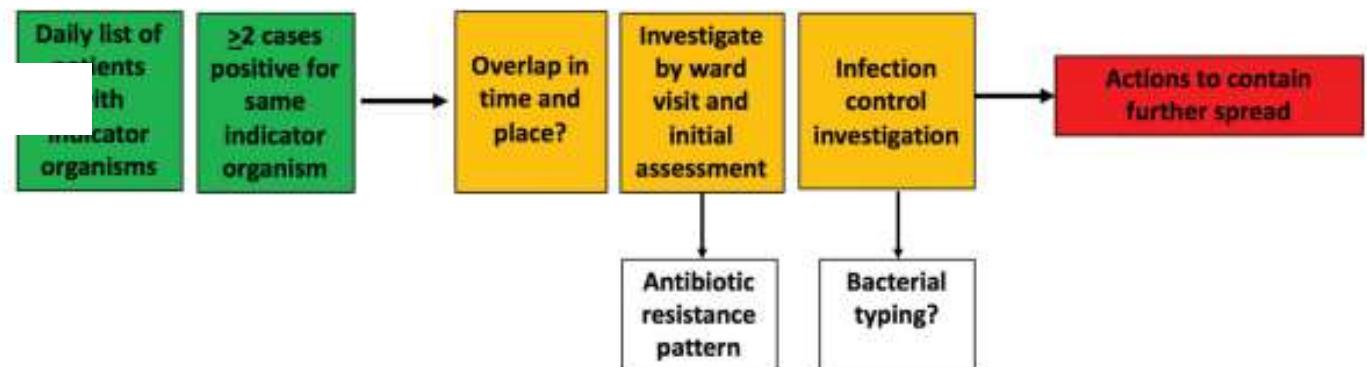
→ Uncovered the previously unrecognized international spread of a near pandrug-resistant nosocomial pathogen, identifiable by a rifampicin-resistant phenotype.

International spread of the epidemic KPC ST258/512 clone



Paradigm shift: real-life example?

NGS-based
outbreak detection



(b)



Peacock et

Fig. 1. Current and proposed approach to the detection of hospital outbreaks. (a) Current practice for the detection of hospital out-

Original Article

Integration of genomic and clinical data augments surveillance of healthcare-acquired infections

Doyle V. Ward PhD^{1,2} , Andrew G. Hoss PhD³, Raivo Kolde PhD³, Helen C. van Aggelen PhD³, Joshua Loving PhD³, Stephen A. Smith BS⁴, Deborah A. Mack RN, CIC⁵, Raja Kathirvel MS⁴, Jeffery A. Halperin MBA⁴, Douglas J. Buell BA⁶, Brian E. Wong MS⁴, Judy L. Ashworth MS⁴, Mary M. Fortunato-Habib BSN, MS, DNP⁴, Liyi Xu PhD³, Bruce A. Barton PhD⁷, Peter Lazar BS⁷, Juan J. Carmona PhD, MPH⁴, Jomol Mathew PhD⁷, Ivan S. Salgo MD⁴, Brian D. Gross MS⁴ and Richard T. Ellison III MD^{2,8}

Objective: To recognize transmission clusters and identify cross-transmission events, using WGS

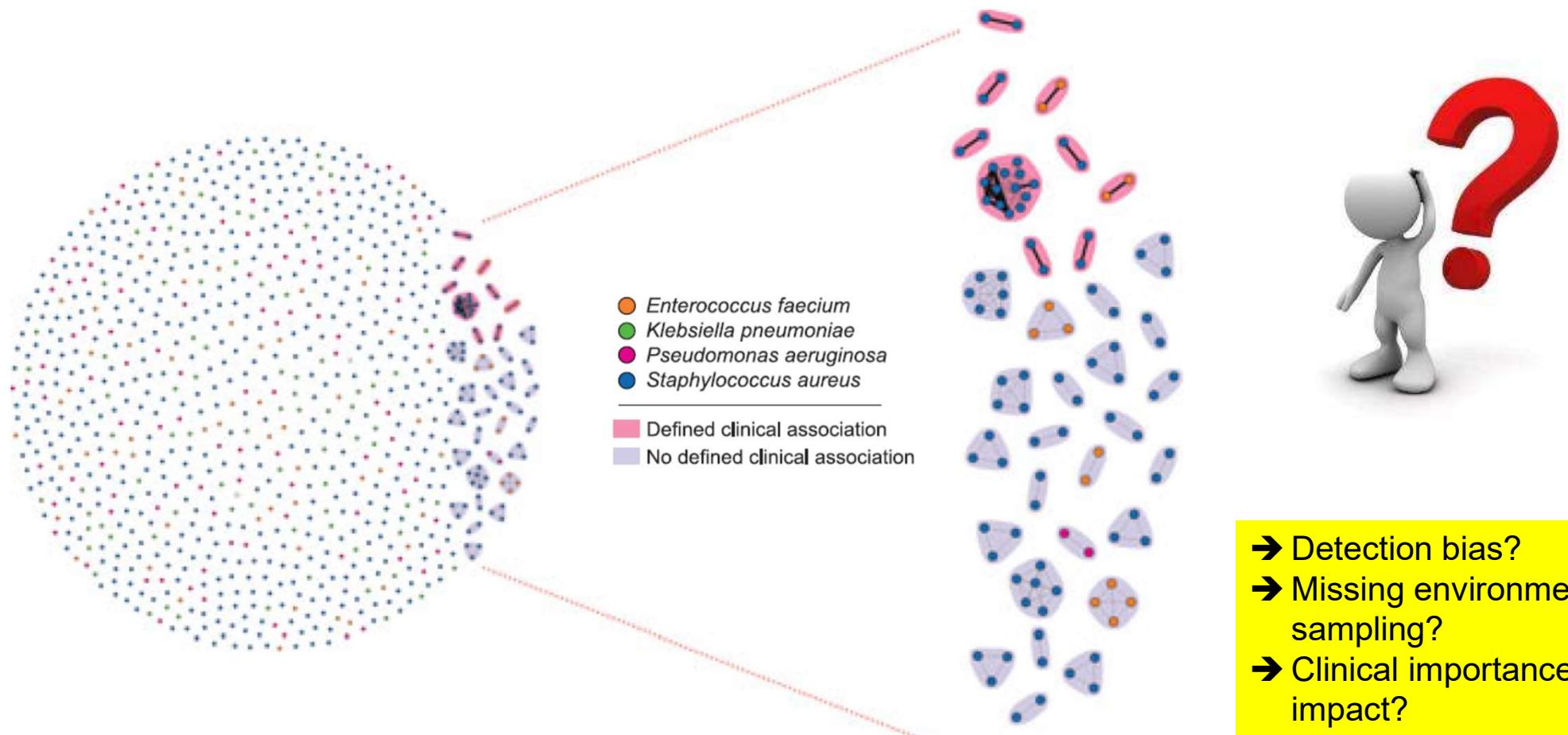
Methods (retrospective cohort study):

- Clinical isolates of *Staphylococcus aureus*, *Enterococcus faecium*, *Pseudomonas aeruginosa*, and *Klebsiella pneumoniae* obtained at UMass from September 1, 2016, to September 30, 2017.
- Isolate genomes were sequenced, followed by single-nucleotide variant analysis
- Use of a cloud-computing platform for WGS analysis and cluster identification

Main results

- Most strains of the 4 studied pathogens were unrelated
→ Endogenous acquisition or outside transmission
- 34 potential transmission clusters involving 96 patients:
 - 25 clusters: no clinical or epidemiological associations
 - 9 clusters: obvious clinical associations
→ only 1 cluster suspected by routine manual surveillance
 - 28 *S. aureus* clusters, 5 *E. faecium* clusters, 1 cluster of *P. aeruginosa*
- Largest cluster: 21 MRSA isolates from 13 patients with community-onset MRSA infections

Identification of suspect transmission events among patients



Ward DV et al, ICHE, 2019; 40: 649-55.

Environmental reservoirs of MDRO

Infection Control & Hospital Epidemiology (2019), **40**, 47–52
doi:10.1017/ice.2018.275



Original Article

A prospective study of transmission of Multidrug-Resistant Organisms (MDROs) between environmental sites and hospitalized patients—the TransFER study

- 44% of hospital rooms contaminated with MDROs after terminal cleaning
- Transfer of bacteria between environment and patient: 18% of admissions
- Occurred early in admission

A *Candida auris* Outbreak and Its Control in an Intensive Care Setting

David W. Eyre, D.Phil., Anna E. Sheppard, Ph.D., Hilary Madder, F.A.N.Z.C.A., Ian Moir, Ruth Moroney, M.Sc., T. Phuong Quan, M.Sc., David Griffiths, B.Sc., Sophie George, M.Sc., Lisa Butcher, M.Sc., Marcus Morgan, M.Sc., Robert Newnham, Mary Sunderland, B.Sc., Tiphanie Clarke, B.A., Dona Foster, Ph.D., Peter Hoffman, B.Sc., Andrew M. Borman, Ph.D., Elizabeth M. Johnson, Ph.D., Ginny Moore, Ph.D., Colin S. Brown, F.R.C.Path., A. Sarah Walker, Ph.D., Tim E.A. Peto, F.R.C.P., Derrick W. Crook, F.R.C.Path., and Katie J.M. Jeffery, Ph.D.

→The transmission of *C. auris* was found to be linked to reusable axillary temperature probes, indicating that this emerging pathogen can persist in the environment and be transmitted in health care settings.



Analytic (sequencing)

Potential biases & challenges during the analyses



Storage parameters:

T°C, time, media, UV, container

Extraction parameters:

enzymatic, mechanical (fav. GPB)

Size selection:

gels (melting~dec. AT-rich sequences)

Library preparation:

PCR approaches (heterogeneous affinities)

The right platform

Illumina

454,
Ion torrent

PacBio,
MinION

Accurate
Takes time(days)

! False positive
variant calling

! False positive
variant calling

	Pre-analytic	Analytic (sequencing)	Analytic (analysis)	Post-analytic
		cgMLST	wgMLST	hqSNP
Discrimination		Alleles in the Core genome	Alleles in the Pan-genome	SNP
Mapping		Manually curated database (beware of new genomes)	Difficult to define alleles, constant evolution	Difficult to create a nomenclature
Inter-facility Comparability		Robust	Allele definition is sensitive to assembler and parameter choices	Sensitive to parameter variations (reference, SNP calling filters, coverage)
Reliability		Set of predefined core genes	Consider accessory genomes (variability, paralogous genes)	Depends of the reference genome

«Clonal or not clonal ? Investigating hospital outbreaks of KPC producing Klebsiella pneumoniae with whole genome sequencing», E.Ruppé, CMI, 2017



Pre-analytic

Method (typing)

Analytic (WGS)

Post-analytic

What is close enough ?

Relatedness thresholds

SNPs: nb of single-nucleotide variant differences

CAVE

- Clock speed is different among pathogens
- Genetic recombination events
- Intra-individual variation

Table 1

Examples of relatedness criteria for wg/cgMLST and SNP typing schemes of representative clinically relevant bacteria

Organism	Relatedness threshold ^a	References
	wg/cgMLST (allele) SNPs	
<i>Acinetobacter baumannii</i>	≤8	[25,26]
<i>Brucella</i> spp.	Epidemiologic validation in progress ^b	http://www.applied-maths.com/applications/wgmlst
<i>Campylobacter coli</i> , <i>C. jejuni</i>	≤14	[27,28]
<i>Cronobacter</i> spp.	Epidemiologic validation in progress ^b	http://www.applied-maths.com/applications/wgmlst
<i>Clostridium difficile</i>	Epidemiologic validation in progress ^b	[29], http://www.cgmlst.org/ncs , http://www.applied-maths.com/applications/wgmlst
<i>Enterococcus faecium</i>	≤20	[30]
<i>Enterococcus raffinosus</i>	Epidemiologic validation in progress ^b	http://www.applied-maths.com/applications/wgmlst
<i>Escherichia coli</i>	≤10	[31,32], https://enterobase.warwick.ac.uk/
<i>Francisella tularensis</i>	≤1	[33,34]
<i>Klebsiella oxytoca</i>	Epidemiologic validation in progress ^b	http://www.applied-maths.com/applications/wgmlst
<i>Klebsiella pneumoniae</i>	≤10	[35,36]
<i>Legionella pneumophila</i>	≤4	[37]
<i>Listeria monocytogenes</i>	≤10	[38,39]
<i>Mycobacterium abscessus</i>	Epidemiologic validation in progress ^b	[40]
<i>Mycobacterium tuberculosis</i>	≤12	[41]
<i>Neisseria gonorrhoeae</i>	Epidemiologic validation in progress ^b	[42], http://www.applied-maths.com/applications/wgmlst
<i>Neisseria meningitidis</i>	Epidemiologic validation in progress ^b	http://www.cgmlst.org/ncs
<i>Pseudomonas aeruginosa</i>	≤14	[31,43]
<i>Salmonella dublin</i>	Epidemiologic validation in progress ^b	[44], https://enterobase.warwick.ac.uk/
<i>Salmonella enterica</i>	Epidemiologic validation in progress ^b	[45], http://www.cgmlst.org/ncs , http://www.applied-maths.com/applications/wgmlst , https://enterobase.warwick.ac.uk/
<i>Salmonella typhimurium</i>	Epidemiologic validation in progress ^b	[46], https://enterobase.warwick.ac.uk/
<i>Staphylococcus aureus</i>	≤24	[47,48]
<i>Streptococcus suis</i>	Epidemiologic validation in progress ^b	[49]
<i>Vibrio parahaemolyticus</i>	≤10	[50]
<i>Yersinia</i> spp.	0	[51]

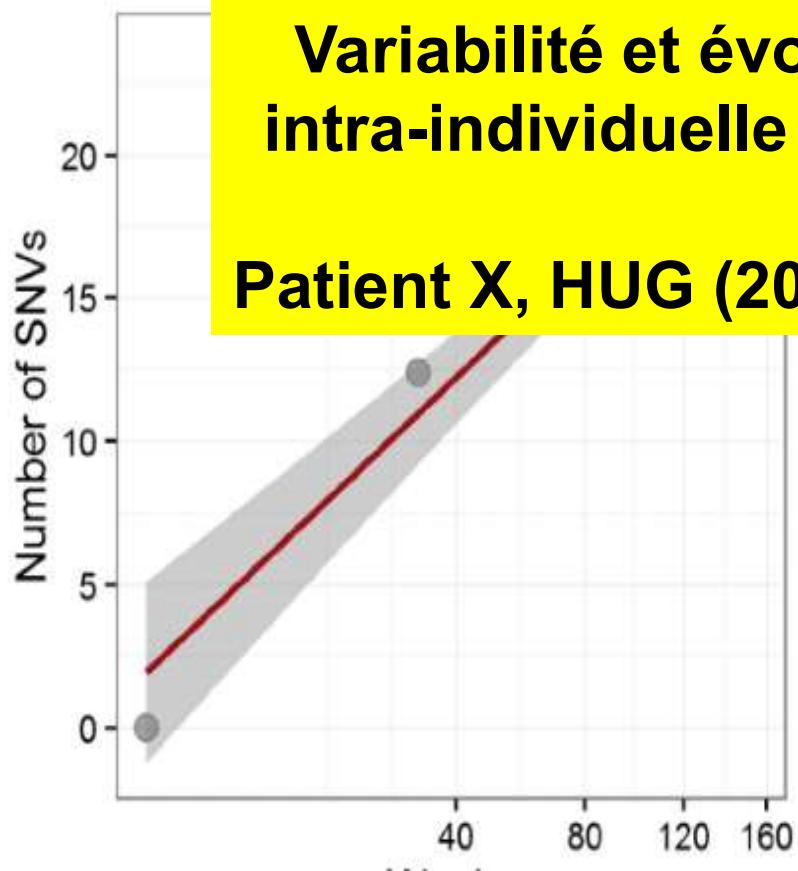
cg, core genome; MLST, multilocus sequence typing; SNP, single nucleotide polymorphism; wg, whole genome.

^a Data often represent single studies that can be used to begin formulation of species-specific interpretation criteria. Thus, these data should be coupled with newly published similar studies to ensure that resulting values are not atypical and can be generally applied.

^b Proposed wg/cgMLST schemes are available online (<http://www.cgmlst.org/ncs>, <http://www.applied-maths.com/applications/wgmlst>, <https://enterobase.warwick.ac.uk/>) but as yet have not been epidemiologically validated.

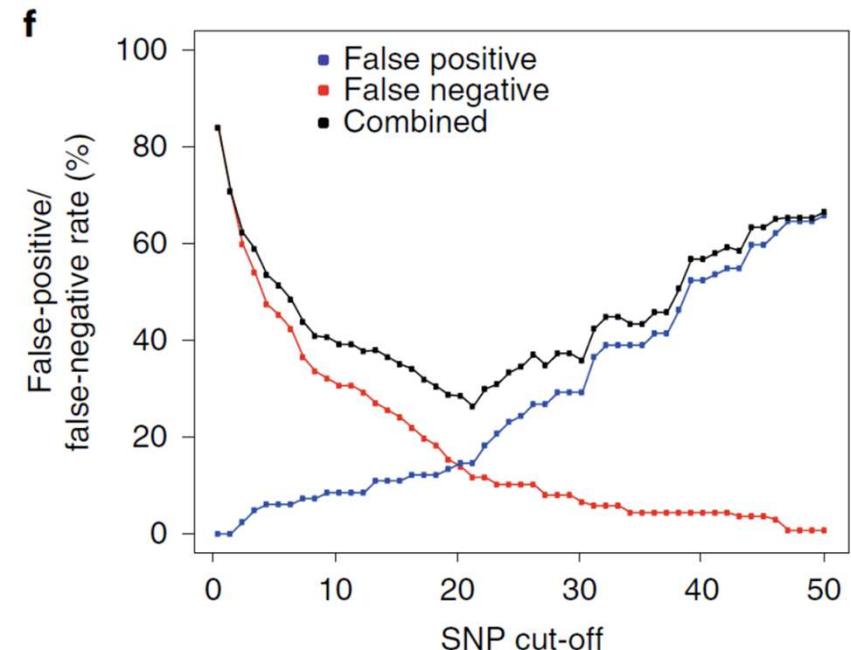
Feedback 

Variabilité et évolution intra-individuelle de KPC Patient X, HUG (2012-2015)



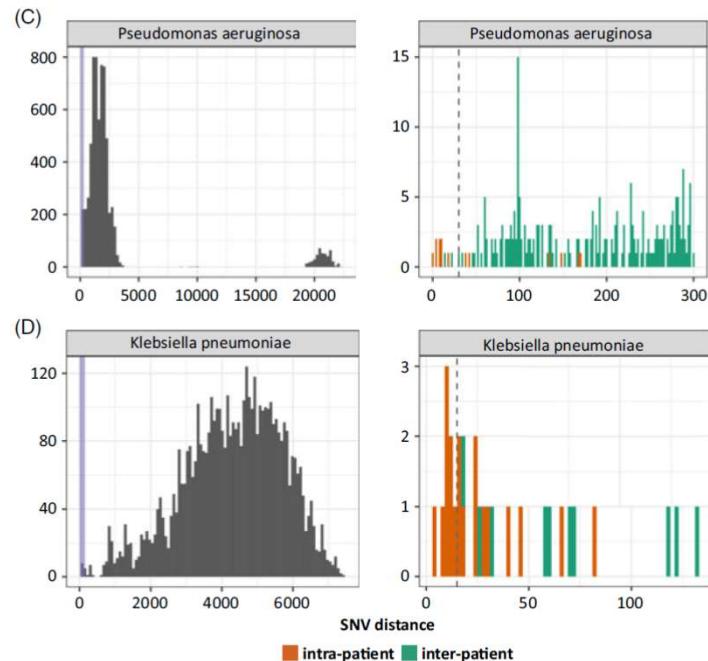
Epidemic of carbapenem-resistant *Klebsiella pneumoniae* in Europe is driven by nosocomial spread

Sophia David¹, Sandra Reuter², Simon R. Harris³, Corinna Glasner⁴, Theresa Feltwell³, Silvia Argimon¹, Khalil Abudahab¹, Richard Goater¹, Tommaso Giani⁵, Giulia Errico⁶, Marianne Aspbury⁷, Sara Sjunnebo⁸, the EuSCAPE Working Group⁹, the ESGEM Study Group¹⁰, Edward J. Feil¹¹, Gian Maria Rossolini^{5,12}, David M. Aanensen^{1,13,14*} and Hajo Grundmann^{1,2,4,14*}



Value of 21 SNPs for discrimination of hospital clusters

Intrapatient versus inter-patient SNP distances to inform thresholds for identification of transmission events



Threshold: 15 SNP for *Klebsiella pneumoniae*

Ward DV et al, ICHE, 2019; 40: 649-55.

Pre-analytic

Method (typing)

Analytic (WGS)

Post-analytic

What is close enough ?

Relatedness thresholds

SNPs: nb of single-nucleotide variant differences

CAVE

THUS

- Clock speed is different among pathogens
 - Genetic recombination events
 - Intra-individual variation
-
- Consider suggested thresholds more as a guideline
 - ➔ Interpret epidemiological links on a case by case basis
 - ➔ Interpret organism by organism (specific population genetics)

«Whole genome sequencing options for bacterial strain typing and epidemiologic analysis based on single nucleotide polymorphism versus gene-by-gene-based approaches», A.C.Schürch, CMI, 2018

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<i>Listeria monocytogenes</i>	≤10	[38,39]
<i>Mycobacterium abscessus</i>	Epidemiologic validation in progress ^b	[40]
<i>Mycobacterium tuberculosis</i>	≤12	[41]
<i>Neisseria gonorrhoeae</i>	Epidemiologic validation in progress ^b	[42], http://www.applied-maths.com/applications/wgmlst
<i>Neisseria meningitidis</i>	Epidemiologic validation in progress ^b	http://www.cgmlst.org/ncs
<i>Pseudomonas aeruginosa</i>	≤14	[31,43]
<i>Salmonella dublin</i>	Epidemiologic validation in progress ^b	[44], https://enterobase.warwick.ac.uk/
<i>Salmonella enterica</i>	Epidemiologic validation in progress ^b	[45], http://www.cgmlst.org/ncs , http://www.applied-maths.com/applications/wgmlst , https://enterobase.warwick.ac.uk/
<i>Salmonella typhimurium</i>	Epidemiologic validation in progress ^b	[46], https://enterobase.warwick.ac.uk/
<i>Staphylococcus aureus</i>	≤24	[47,48]
<i>Streptococcus suis</i>	Epidemiologic validation in progress ^b	[49]
<i>Vibrio parahaemolyticus</i>	≤10	[50]
<i>Yersinia</i> spp.	0	[51]

cg, core genome; MLST, multilocus sequence typing; SNP, single nucleotide polymorphism; wg, whole genome.

^a Data often represent single studies that can be used to begin formulation of species-specific interpretation criteria. Thus, these data should be coupled with newly published similar studies to ensure that resulting values are not atypical and can be generally applied.

^b Proposed wg/cgMLST schemes are available online (<http://www.cgmlst.org/ncs>, <http://www.applied-maths.com/applications/wgmlst>, <https://enterobase.warwick.ac.uk/>) but as yet have not been epidemiologically validated.

Feedback 

Eclosion de Klebsielles multi-R aux HUG en 2015

Cas index



Cas secondaire N° 1



Cas secondaire N° 2



	R	Amoxicilline	
256	R	Co-amoxiclav	
	R	Piperacilline	
	R	Piperac.+tazob	256 256
	R	Cefoxitine	
	R	Cefuroxime	
	R	Ceftazidime	
	R	Ceftriaxone	
	R	Cefepime	96 96
	R	Imipenem	>32 >32
	R	Meropeneme	>32 >32
	R	Ertapénem	32 32
	R	Aztreonam	
	R	Amikacine	2 2
	S	Gentamicine	
	R	Norfloxacine	
	R	Ciprofloxacine	
	R	Co-trimoxazole	48 48
	S	Fosfomycine	2 2
	I	Tigécycline	32 32
	R	Furanes	
	S R	0.750.750	

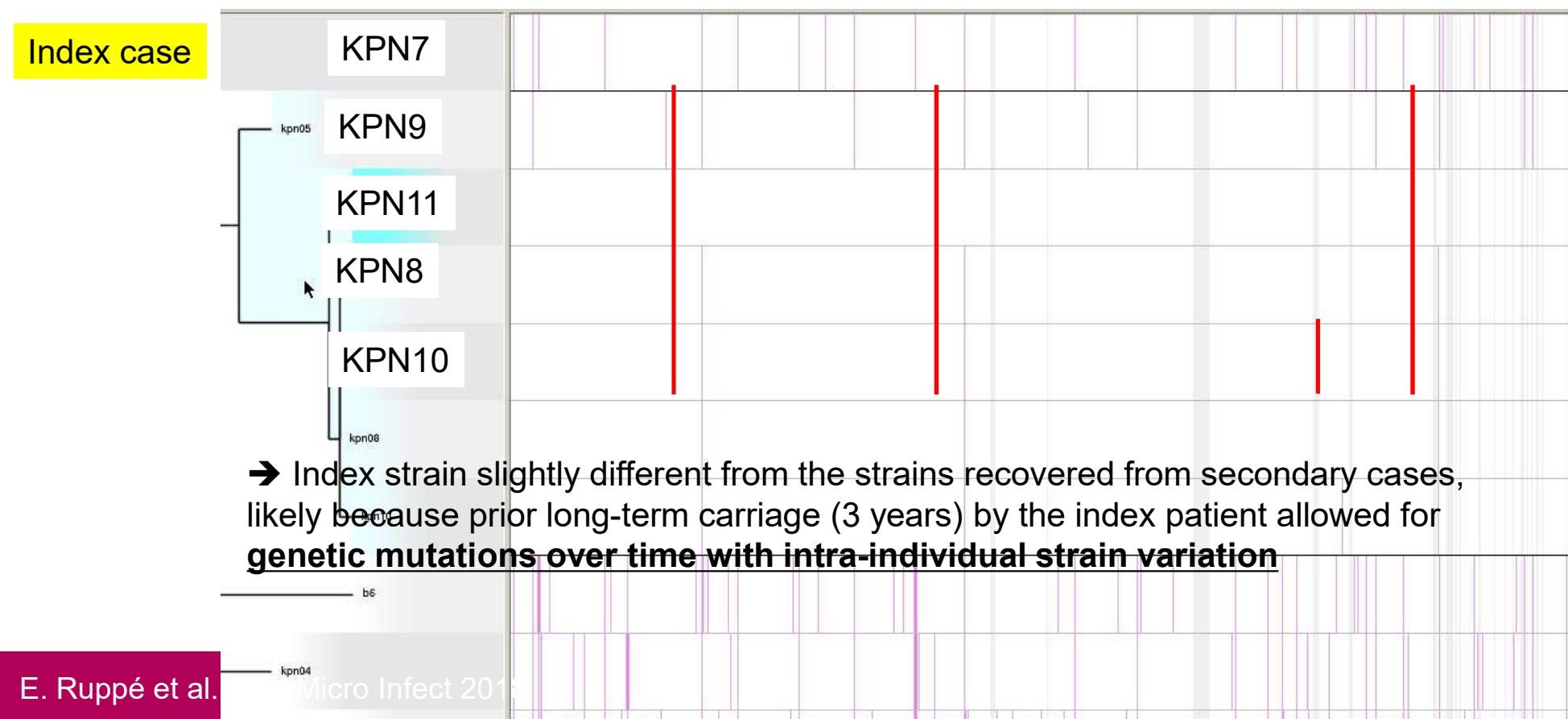


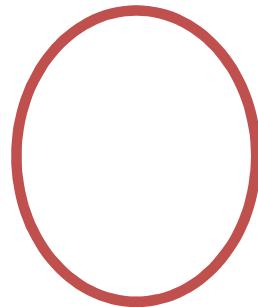
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SNVs analysis from whole common DNA

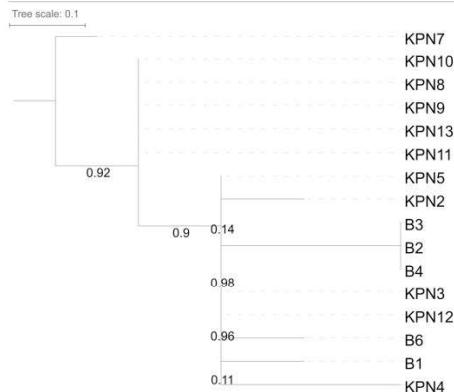
Whole genome analysis: KPN7 had 3 SNV differences with others, KPN10 has another SNV





Blinded assessment of clonal relationships by wgMLST (634 genes)

Fifteen of the 18 strains belonged to ST512, while isolates KPN1, B5 and B6 belonged to ST258. From the wgMLST analysis, we first observed that KPN7 was distinct from other strains and had one SNV difference with the closest cluster that included five strains (KPN8, -9, -10, -11 and -13) (Fig. 2, Table 1). In addition, another cluster of three strains was observed (B2, B3 and B4). The other strains appeared to be unrelated at this level of analysis, with KPN1 and B5 being the most distant strains from KPN7 (Table 1); their involvement in any outbreak caused by KPN7 was thus unlikely.



[Download high-res image \(170KB\)](#) [Download full-size image](#)

Fig. 2. Phylogenetic tree with bootstrap values obtained by comparison of genes included in *Klebsiella pneumoniae* wgMLST. KPN7 is suspected index strain for outbreak. B5 and KPN1 do not appear on tree because of their high distance. wgMLST, whole-genome multilocus sequence typing

Scarthorn

SNP and ARGs (accessory genome)

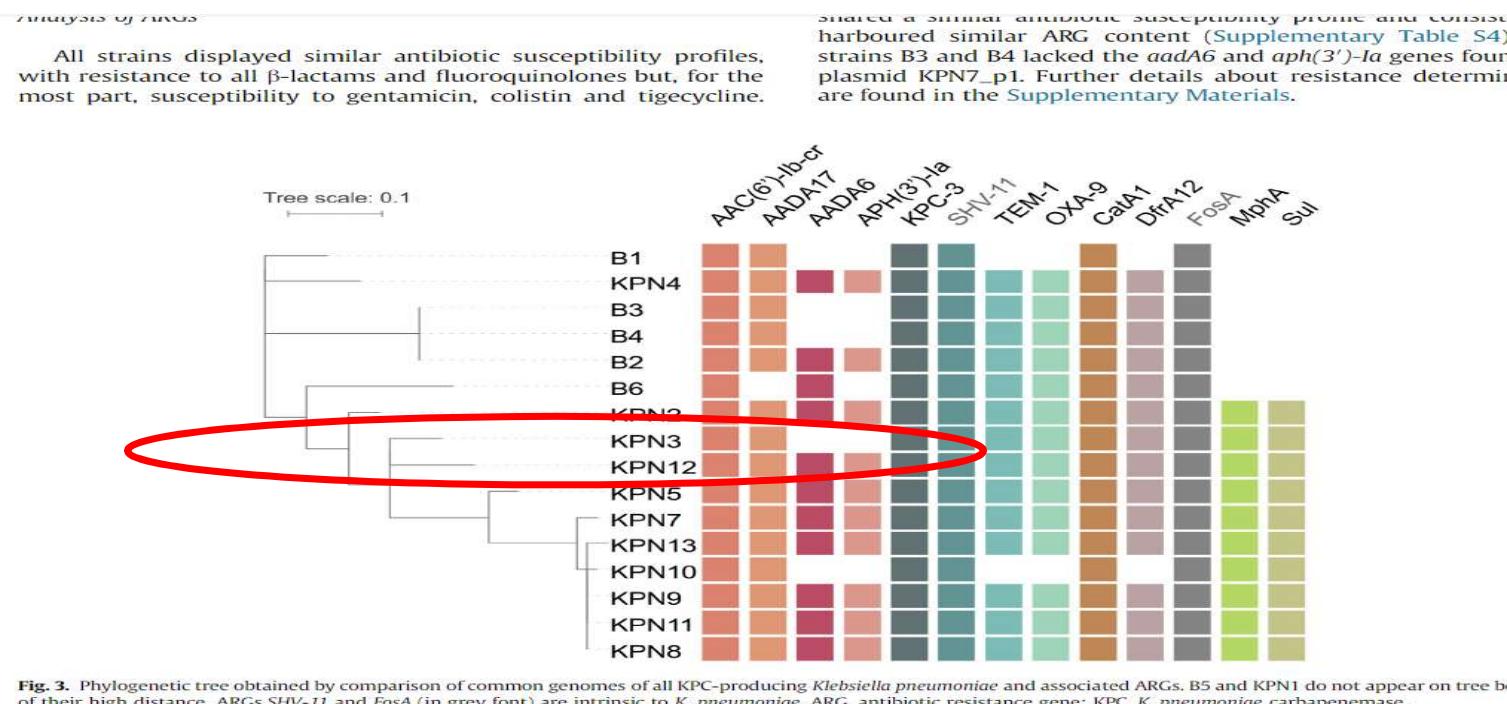


Fig. 3. Phylogenetic tree obtained by comparison of common genomes of all KPC-producing *Klebsiella pneumoniae* and associated ARGs. B5 and KPN1 do not appear on tree because of their high distance. ARGs SHV-11 and FosA (in grey font) are intrinsic to *K. pneumoniae*. ARG, antibiotic resistance gene; KPC, *K. pneumoniae* carbapenemase.



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2 outbreak investigations from Geneva

MRSA

CPE



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Friday, June 30

Phone rings:

„Hello...! We have a problem down here at the NICU
– can you help us ?“



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"Do you want to speak to the man
in charge or the nurse who knows
what's going on?"

Veni vidi ... (vici ?)

Julius Caesar, De Bello Gallico

You go in there and have a look...

- **What?** • MRSA outbreak
- **Who?** • 11 neonates, 2 mothers
- **Where?** • NICU & nursery
- **When?** • Over the last 3 months
- **Severe?** • No – mostly carriage



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L'ESSENTIEL, C'EST VOUS.

What would you recommend for the next week – priority action items except?

- A. Isolate or cohort MRSA carriers
- B. Reinforce hand hygiene
- C. Screening of all hospitalized neonates
- D. Implement active MRSA surveillance for all new admissions
- E. Molecular typing of MRSA isolates

What would you recommend for the next week – priority action items except?

- A. Isolate or cohort MRSA carriers
- B. Reinforce hand hygiene
- C. Screening of all hospitalized neonates
- D. Implement active MRSA surveillance for all new admissions
- E. Molecular typing of MRSA isolates**

Epidemic curve MRSA outbreak, NICU (HUG)

8

7

6

5

4

3

2

1

1999 2000

ANTIBIOTIQUES (1)x1 SCN. (2)x3 MRSAe Interpré. Interpré.

Penicilline G	RESIST	RESIST
Fluclouxacilline	RESIST	RESIST
Amikacine	RESIST	RESIST
Gentamicine	S	RESIST
Norfloxacine	RESIST	RESIST
Ciprofloxacine	RESIST	RESIST
Clindamycine	S	RESIST
Erythromycine	RESIST	RESIST
Acide fusidique	INTERM	S
Co-trimoxazole	RESIST	S
Fosfomycine		S
Rifampicine	S	S
Vancomycine	S	S
Teicoplanine	S	S

Document 75% du dossier Observations Infirmières Visualiseur de documents Nouveaux documents

25-07-2000 11:00:00 EXAMEN DE L'URINE

H.U.G.: Laboratoire Central de Bactériologie (Document DEFINITIF si...)

MEOMA+ 1b 16/U//UU U8HUU trottis inguinal
MEOMA+ 18 laboratoire Central de Bactériologie WALKOWSKI Jason, typ Page 4
1W/U//UU 12HUS urine poche a urines Mercredi 04/08/2000 15h56

MEOMA+ 25 U8HUU trottis inguinal 16 UU25
25/U//UU 10HBS trottis de nez 18 U145
MEOMA+ 25 12HUS urine poche a urines
MEOMA+ 25/U//UU 10HBS trottis inguinal 25 U130
MEOMA+ 25/U//UU 11HUU trottis umbilic 25 U151
10HBS trottis inguinal 25 U155
11HUU trottis umbilic

QUANTIFICATION: le chiffre (1UE4 :
ABS = absence de, PRES = présence
++ = nombreux, +++ = très nombreux)

ANTIBIOTIQUE (1)x6 MRSAe Interpré.

Penicilline G	RESIST	RESIST
Fluclouxacilline	RESIST	RESIST
Amikacine	S	S
Gentamicine	S	S
Norfloxacine	S	S
Ciprofloxacine	S	S
Clindamycine	S	S
Erythromycine	S	S
Acide fusidique	RESIST	RESIST
Co-trimoxazole	S	S
Fosfomycine	S	S
Rifampicine	S	S
Vancomycine	S	S
Teicoplanine	S	S

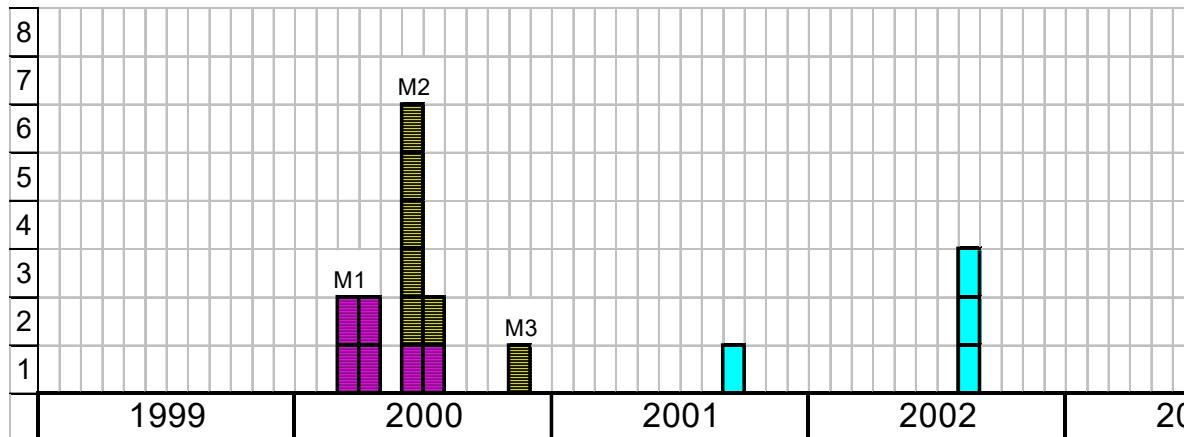
INTERPRETATION: le chiffre (1UE4 = 1U/100 etc.) indique le nombre de gérres par ml de prélevement.
S = sensibilité, R = résistance, N = non-interprétable, MEG = négatif, POS = positif,
++ = nombreux, +++ = très nombreux.

(Cx n = Référence à l'antibiogramme
RESIST ou R = Résistant S = Sensible
N = Non-interprétable)

Veuillez vous référer au Répertoire des substances actives et aux notices de sous-traitance. Ce rapport ne peut être utilisé sans l'autorisation de l'Institut de Santé publique et de la Sécurité sociale.

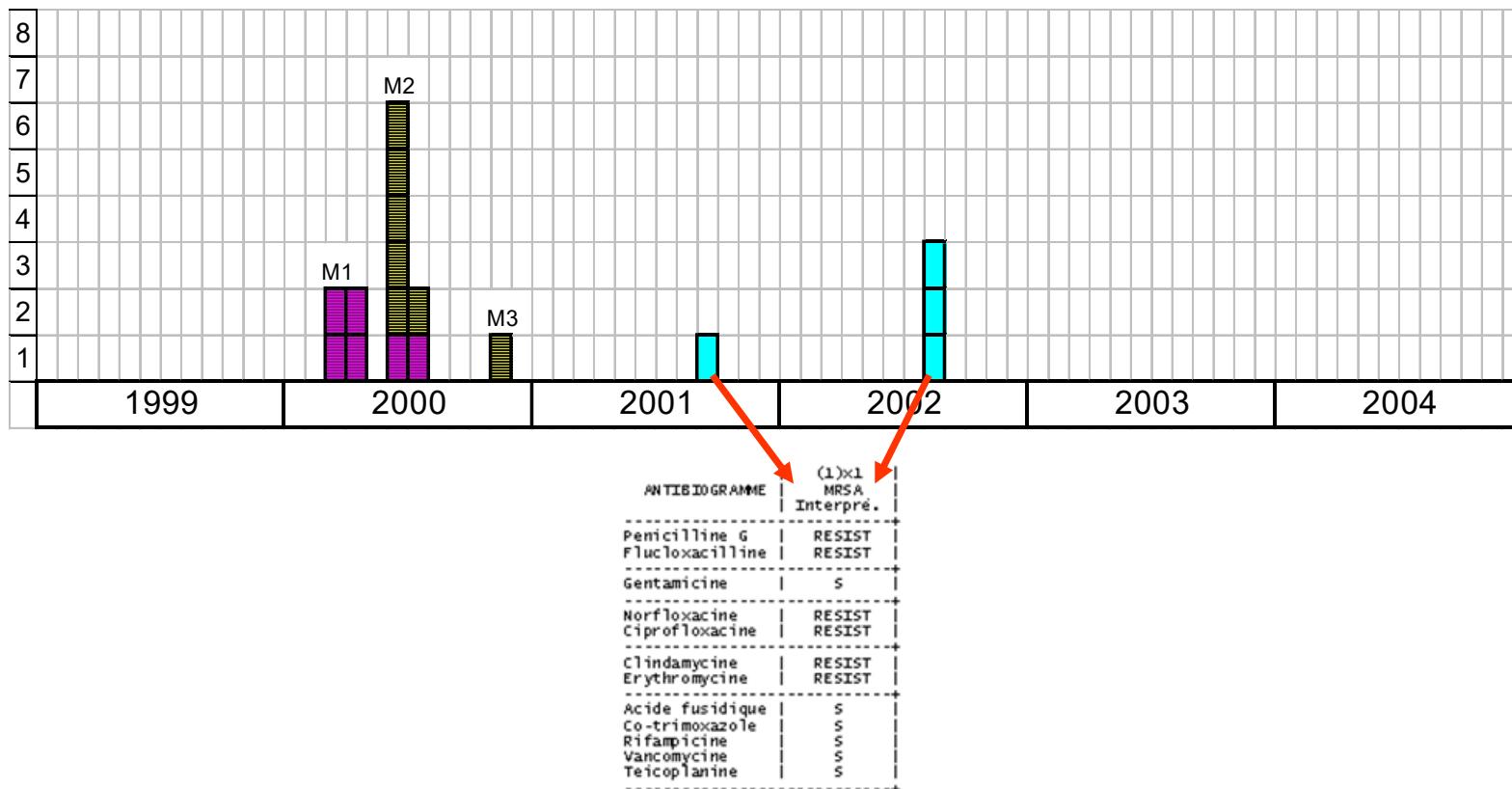
Sax et al. J Hosp Infect 2006; 63: 93-100

Control measures MRSA outbreak, NICU (HUG)



- A. Isolate or cohort MRSA carriers
- B. Reinforce hand hygiene
- C. Screening of all neonates & mothers
- D. Implement active MRSA surveillance (admission & discharge)

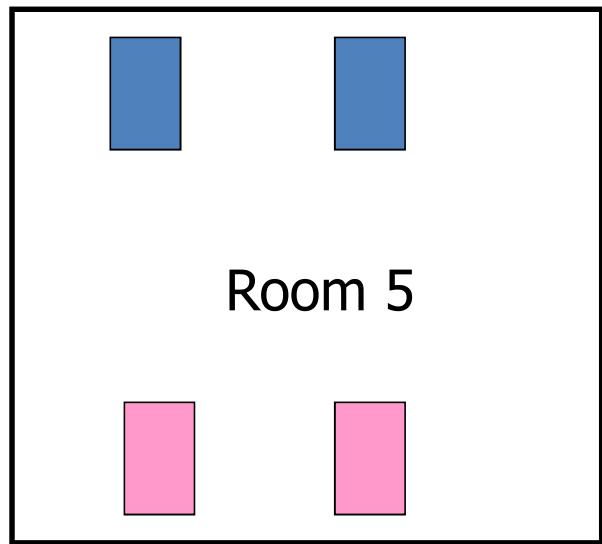
Epidemic curve MRSA outbreak, NICU (HUG)



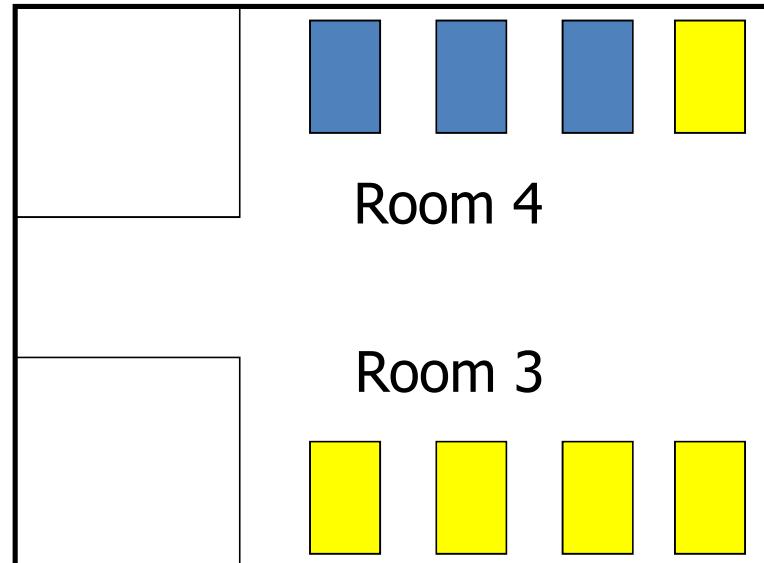
Sax et al. J Hosp Infect 2006; 63: 93-100

NICU at HUG

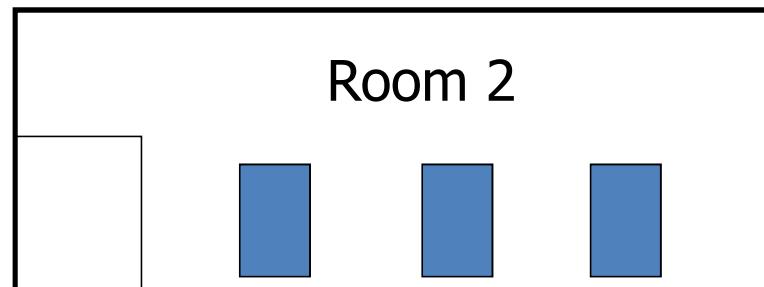
June/July 2000



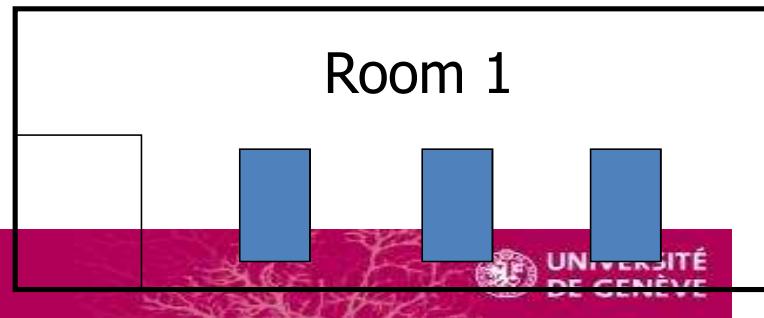
Room 5



Room 4



Room 2



Room 1

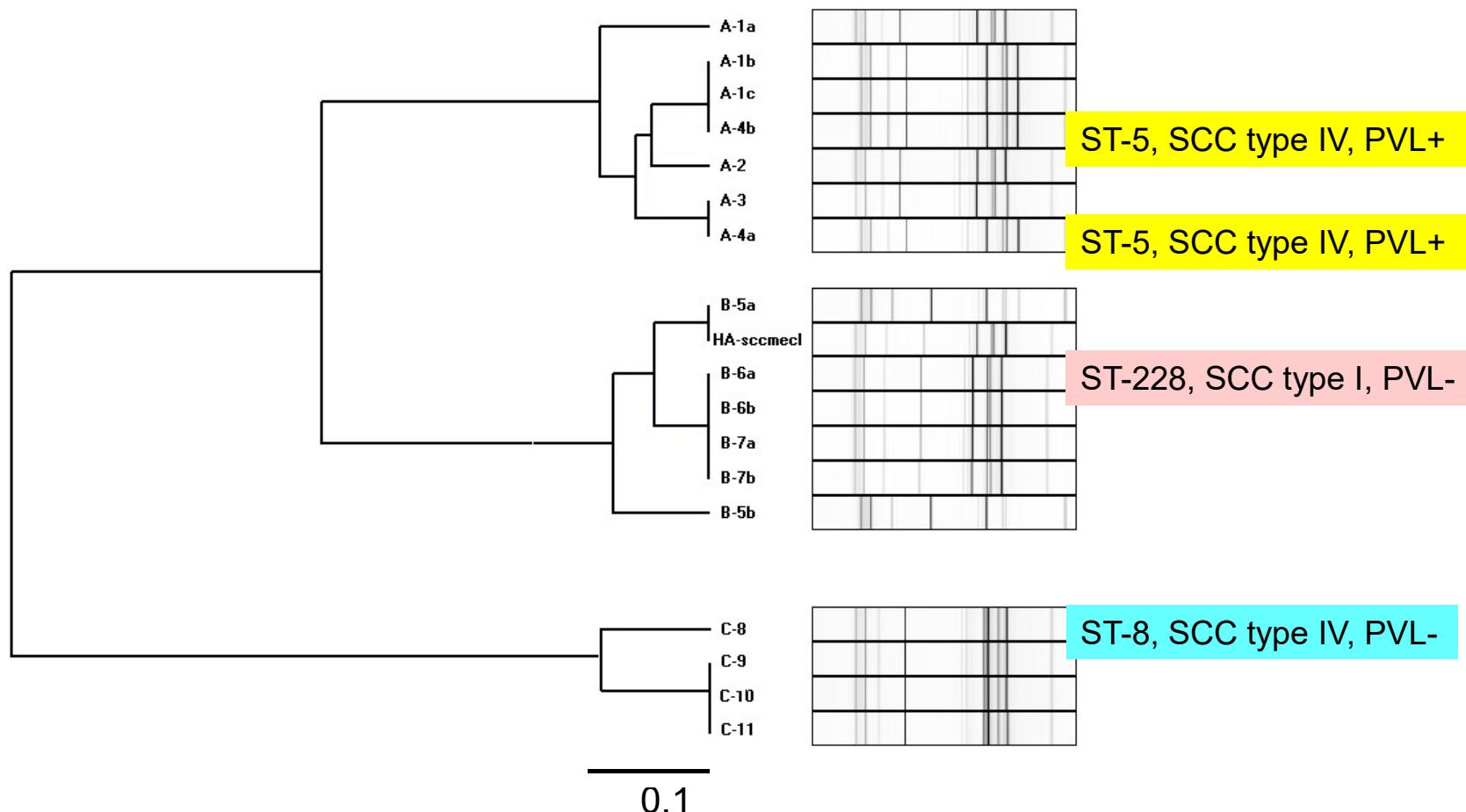
Legend

PVL+ CA-MRSA strain

Multi-R strain (endemic at HUG)

Sax et al. J Hosp Infect 2006; 63: 93-100

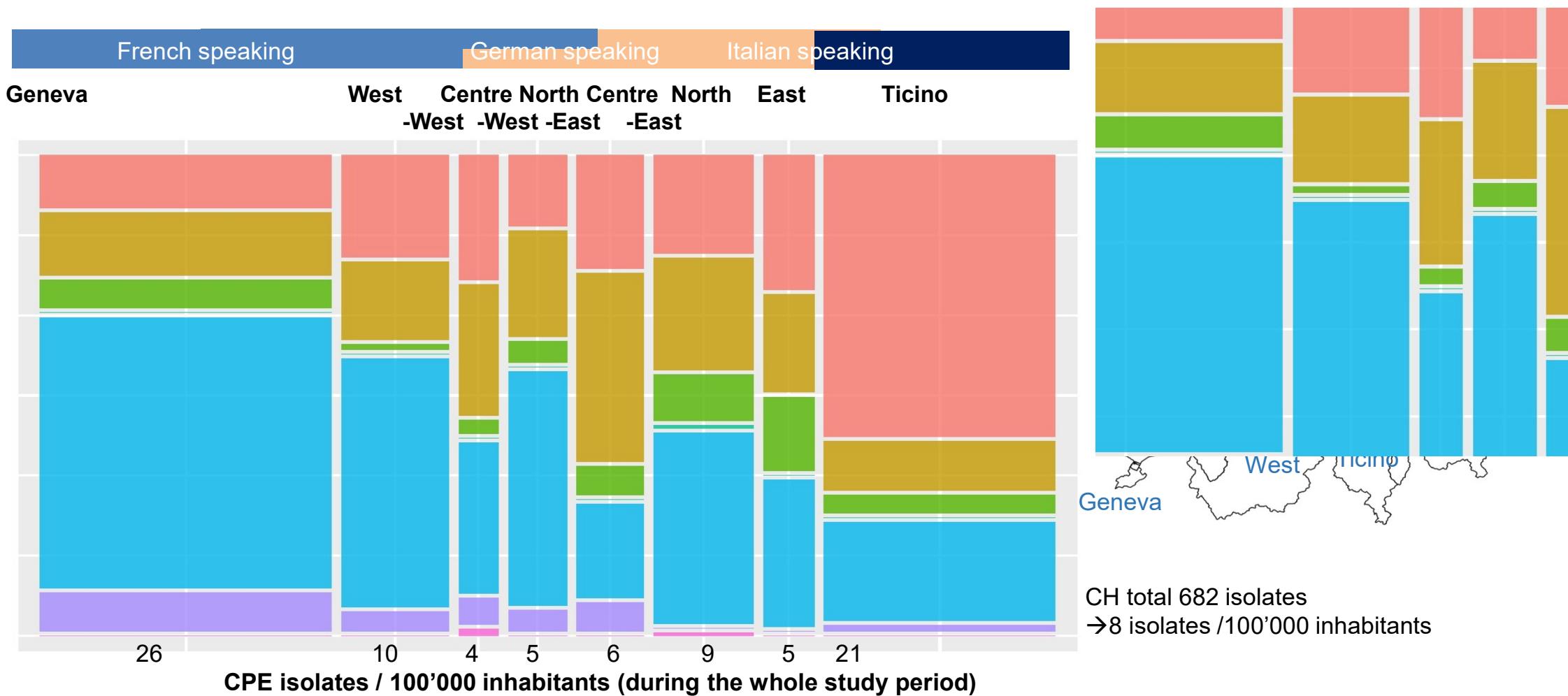
Molecular Epidemiology



Orthopedic ward, Geneva

OXA-181 cluster, summer 2019

CPE isolates in CH 2013 - 2018



Courtesy: A. Kronenberg (anresis.ch)



1^{ère} investigation

04-06.19



18.06.2019 – 5AL (orthopédie septique)

Départ d'un patient porteur (M.H.) de *K.pneumoniae* KPC

Hospitalisé au 5AL du 22.05-18.06.19

→ Dépistage de l'unité

Détection fortuite de 3 porteurs de carbapénémases (OXA-181)

(M.K.) *Citrobacter* sp BLSE & OXA-181 (FA) – au 5AL du 28.05-25.06.19

(M.P.) *Citrobacter* sp BLSE & OXA-181 (FA) – au 5AL du 12.04-19.06.19

(M.B.) *Citrobacter* sp BLSE & OXA-181 (FA) – au 5AL du 28.05-03.07.19



1ère investigation

03.2019-04-06.19



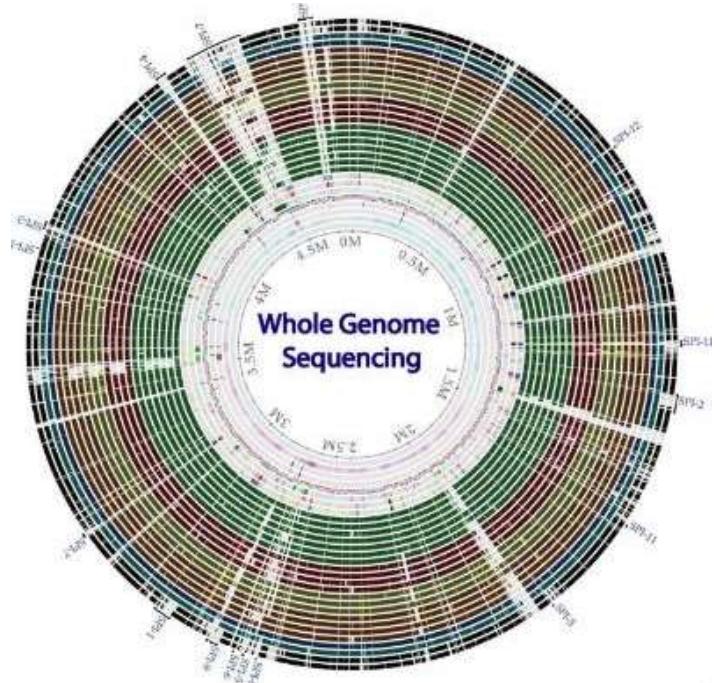
Recherche d'un cas index

(M.R.) *Klebsiella pneumoniae* OXA-181 – au 5AL du **08.03-20.03.19**

- Originaire du Sri Lanka & hospitalisation en Inde (27.02-06.03.19)
- Mesures CONTACT PLUS durant son hospitalisation et son suivi
- Bonne compliance de l'équipe, mauvaise compliance du patient

MR2

Possible transfert horizontal inter-espèce du gène OXA-181 ?



Diapositive 61

MR2

MARTISCHANG Romain; 17/10/2019

1ère investigation

03.2019 04-06.19

Mais...

Citrobacter

Klebsiella

Profil plasmidique	GRL-89	GRL-90	GRL-91	GRL-92
Col440I				
ColKP3				100
ColRNAI				
IncX3	100	100	100	
IncFIA				99.74
IncFIB (AP001918)				
IncFII (29)				
IncFII (pAMA1167-NDM-5)				98.08
IncY				
IncFII				
ColpVC				
IncFIB (pB171)	100	100	100	
IncQ1				100
IncFIB (pQil)				



Hypothèse épidémiologique infirmée (autre sous-type de OXA: gènes OXA-1 et OXA-232)

2ème investigation

03.2019 04-06.19



Nouveau patient index hypothétique

(M.P.) *Citrobacter* sp BLSE & OXA-181 (FA) – au 5AL du **12.04-19.06.19**

- Circulation libre dans l'ensemble du 5AL
- Non compliance aux mesures de précaution (pt toxicomane)
- Source de contamination primaire par OXA-181 obscure

Et 1 mois plus tard...

2ème investigation

03.2019 04-06.19 07.19



27.08.2019 – 5AL : enquête de prévalence hebdomadaire

(M.N.) *E.coli* BLSE & OXA-181 (FA) – au 5AL du **22.07-30.08.19**

- Multiples opérations (23.07, 31.07, 19.08) aux HUG
- 5 frottis anaux negatifs pour CPE (22.07, 30.07, 16.08, 13.08, 20.08)
- **SANS contact direct avec les 3 autres cas OXA-181 de juin 2019**
- Séquençage à tout hasard...

2^{ème} investigation

Et...

Citrobacter sp BLSE & OXA-181
21.06.2019, 5-AL

E.coli BLSE & OXA-181
27.08.2019, 5AL

	Citrobacter	Klebsiella pneumoniae		Escherichia coli		Ec-101			
		GRL-89	GRL-90	GRL-91	GRL-92	GRL-93	GRL-94	GRL-95	GRL-96
Beta-lactam	blaSHV-33				100				
Beta-lactam	blaOXA-181	100	100	100		100	100	100	100
Beta-lactam	blaCTX-M-15	100	100	100	100		100	100	100
Beta-lactam	blaTEM-1B	100	100	100	100		100	100	100
Beta-lactam	blaOXA-1				100				
Beta-lactam	blaOXA-232				100				100

	Position dans la référence				
GRL-89 Citrobacter sp. 2.6.11.28		14183-14454			20364-20989
GRL-90 Citrobacter sp. 2.6.11.27		14183-14454			20364-20988
Ec-101 Escherichia coli 2.6.16.13		14183-14454			20364-20988
GRL-95 Escherichia coli 2.3.16.21					20380-20970
GRL-91 Citrobacter sp. 2.6.11.26	13221-14457				20364-20988

2^{ème} investigation

Et...

***Citrobacter* sp BLSE & OXA-181**
21.06.2019, 5-AL

	Citrobacter	Klebsiella pneumoniae			Escherichia coli					
		GRL-89	GRL-90	GRL-91	GRL-92	GRL-93	GRL-94	GRL-95	GRL-96	Ec-101
Beta-lactam	blaSHV-33				100					
Beta-lactam	blaOXA-181	100	100	100		100	100	100	100	100
Beta-lactam	blaCTX-M-15	100	100	100	100		100	100	100	100
Beta-lactam	blaTEM-1B	100	100	100	100		100	100	100	
Beta-lactam	blaOXA-1				100					
Beta-lactam	blaOXA-232				100					100

***E.coli* BLSE & OXA-181**
27.08.2019, 5AL

Conclusions hors du domaine accrédité

Nos résultats suggèrent la possibilité que le même plasmide a été trouvé dans les souches :

GRL-89 Citrobacter sp. 2.6.11.28
GRL-90 Citrobacter sp. 2.6.11.27
Ec-101 Escherichia coli 2.6.16.13

Les autres souches présentent un profil unique de délétions par rapport à la référence choisie.

Le séquençage à longs reads (Nanopore ou PacBio) pourrait confirmer nos hypothèses.

Hypothèse épidémiologique probablement confirmée

En attente d'une analyse plasmidique plus détaillée pour confirmer ces résultats

2ème investigation



Hypothèses :

- Réservoir plasmidique environnemental
- Non détection d'un faible inoculum de *Citrobacter* OXA-181 (transfert intra-host)

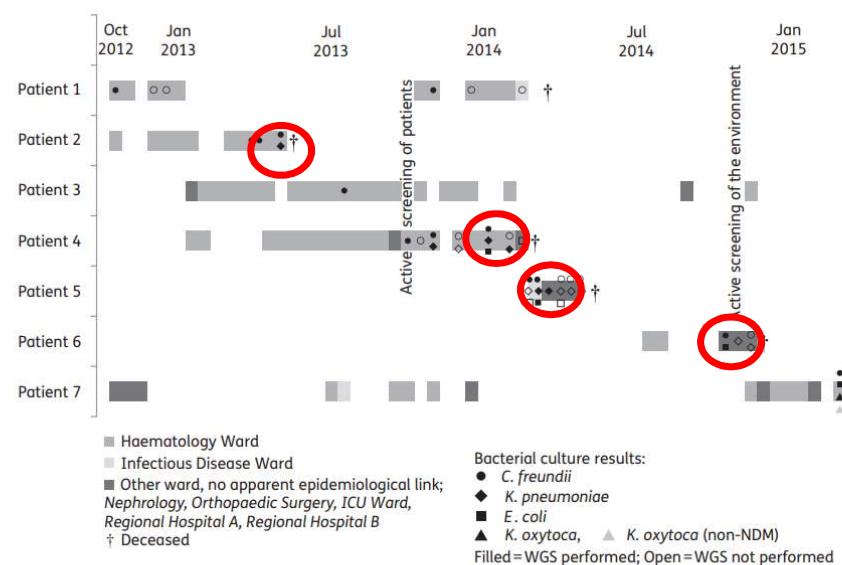


Figure 1. Timeline representing the NDM-1 *C. freundii* outbreak. Ward locations are marked in different shades of grey. Epidemiological links were detected for Patients 1, 2, 3, 4, 6 and 7 in the haematology ward and for Patients 1 and 5 in the infectious disease ward. Screening of all admitted patients and screening of the environment in the haematology ward were performed in October 2013 and November 2014, respectively, without identifying the source of the outbreak.

J Antimicrob Chemother 2016; **71**: 3117–3124
doi:10.1093/jac/dkw289 Advance Access publication 1 August 2016

**Journal of
Antimicrobial
Chemotherapy**

Use of WGS data for investigation of a long-term NDM-1-producing *Citrobacter freundii* outbreak and secondary *in vivo* spread of *bla*_{NDM-1} to *Escherichia coli*, *Klebsiella pneumoniae* and *Klebsiella oxytoca*

Anette M. Hammerum^{1*}, Frank Hansen¹, Hans Linde Nielsen², Lotte Jakobsen¹, Marc Stegger^{1,3}, Paal S. Andersen^{1,3,4}, Paw Jensen⁵, Tue Kjærgaard Nielsen⁶, Lars Hestbjerg Hansen⁶, Henrik Hasman¹ and David Fuglsang-Damgaard²

¹Department of Microbiology and Infection Control, Statens Serum Institut, Copenhagen, Denmark; ²Department of Clinical Microbiology, Aalborg University Hospital, Aalborg, Denmark; ³Pathogen Genomics Division, Translational Genomics Research Institute (TGen), Flagstaff, AZ, USA; ⁴Veterinary Disease Biology, University of Copenhagen, Copenhagen, Denmark; ⁵Department of Haematology, Aalborg University Hospital, Aalborg, Denmark; ⁶Department of Environmental Science, Aarhus University, Roskilde, Denmark

- Denmark: outbreak with *C.freundii* NDM-1
- Investigated by WGS
- Multiple horizontal transfers to *E.coli*, *K.pneumoniae* and *K. oxytoca*

Prise en charge SPCI



1ère investigation 18.06.2019 – 5AL

- Dépistage de l'unité
- Enquêtes de prévalence hebdomadaire (en plus du dépistage à l'admission en routine)
- Recherche, marquage, et dépistage à l'admission des patients (n=59) ayant croisé le premier cas index présumé
- Isolation et cohortage des patients positifs OXA-181 dans l'unité 5-AL, avec aide soignant dédié
- Nettoyage quotidien d'unité et de la salle de traitement au Tristel Fuse Surfaces (22.06.19-fin Sept.)

2ème investigation 18.06.2019 – 5AL

- Recherche, marquage, et dépistage à l'admission des patients (n=121) ayant croisé le deuxième cas index présumé

Conclusions

- La flambée de *Citrobacter* sp producteur de carbapénémase (OXA-181) dans l'unité du 5-AL, département de chirurgie aux HUG est à notre connaissance une première en Suisse.
- La documentation d'une transmission horizontale nosocomiale et inter-espèce du plasmide contenant le gène OXA-181 est aussi une première en Suisse.
- Bonne compliance des soignants aux mesures de précaution
- Mauvaise compliance des patients aux mesures de précaution
- Certaines limitations sont soulignées
 - Limitations dues aux soins (unité d'orthopédie septique)
 - Transmission potentiellement augmentée (écoulements biologiques)
 - Haute densité de consommation antibiotique
 - Limitations environnementales
 - Circulation des patients et leurs familles (suivi ambulatoire en salle de traitement)
 - Absence de local dédié aux patients avec portage BMR
 - Aucun sanitaire dédié à chaque patient (ni salle de bain, ni toilettes)

Propositions SPCI

Au regard des patients à risque

- Une chambre individuelle avec sanitaire dédié pour tout patient
- Des chambres dédiées aux Mesures Spécifiques
- Une salle de traitement dédiée (avec salle d'attente à distance de l'unité)
- Un système de sécurité pour prévenir de la non compliance au respect des mesures mise en place.

Au regard du risque de futures épidémies BMR

- Maintien du dépistage BMR complet par frottis pour tout patient admis
- Maintien de l'enquête de dépistage hebdomadaire au-delà du 30 Sept 2019
- Dépister d'unité lors du départ d'un patient identifié comme porteur de MDRO même si ce patient a été hospitalisé en chambre individuelle
- Activation d'une alerte qui identifie en cas de réadmission les patients à risque d'avoir été contaminés lors d'un séjour hospitalier antérieur par un patient porteur d'une bactérie hautement multi-résistante
- De regrouper les patients en leur dédiant du personnel nécessaire en cas d'épidémie.

Surgical ward, Geneva

NDM cluster 2018-2019

BACKGROUND

From July 2018 to May 2019

3 cases positive for NDM-producing *E.coli* and attributable to our hospital

Fortuitously detected

- 1 case by routine weekly screening in the ICU
- 1 case by screening b/o additional MDRO carriage
- 1 case by urine culture in LTCF after extended hospital in a private room

No foreign travel in the past 12 months

No known risk factor for acquisition of MDRO carriage



Objective: To report an outbreak investigation guided through molecular methods

BACKGROUND

Cas index: Mr. G.

52 years, from United Arab Emirates

Known for inflammatory bowel disease with multiple surgical interventions

- New York (April 2017)
- Geneva (December 2017)

Transfer to Geneva University Hospitals with subsequent ICU stay (25.12.17) → NDM *E.coli* pos

Rehospitalized at HUG in July 2018

Pre-emptive contact precautions



ROUGE-B	M.171225.0128/1	ABS Klebsiella pneumoniae carbapenemase (KPC), par PCR	ABS Acinetobacter sp. multi-résistant	T
25/12/17 07:45	Frottis anal	PRES ^{na} New Delhi metallo-beta-lactamase (NDM), par PCR	(a) +++ Escherichia coli	(1) T
		ABS ^{na} Oxacillinase 48 (OXA-48), par PCR	ABS Enterococcus spp. vancomycine résistant	T
		ABS ^{na} Oxacillinase 181 (OXA-181), par PCR		T
		ABS ^{na} SPM-type carbapenemase, par PCR		T
		ABS ^{na} Verona integron-encoded metallo-beta-lactamase (VIM), par PCR		T
		ABS ^{na} Metallo-beta-lactamase IMP, par PCR		T
		ABS ^{na} GES-type carbapenemase, par PCR		T
		ABS ^{na} SME-type carbapenemase, par PCR		T

antibiogramme (CMI exprimée en mg/L)	
R	Amoxicilline AMOXIC
R	Co-amoxiclav AM.CLA
R	Piperacilline PIPERA
R	Piperac.+tazob PI+TAZ
R	Cefuroxime CEFURO
R	Ceftazidime CEFTAZ
R	Ceftolozane+tazobactam CETOZA
R	Ceftriaxone CEFTRI
R	Cefepime CEEPEPI
I	Imipenem IMIPEN
I	Meropenème MEROPÉ
R	Ertapènem ERTAPE
R	Aztreonam AZTREO
R	Amikacine AMIKAC
R	Gentamicine GENTAM
R	Norfloxacine NORFLO
R	Ciprofloxacine CIPROF
R	Co-trimoxazole COTRIM
S	Fosfomycine FOSFOM
S	Tigécycline TIGECY
S	Furanes FURANE
S	Colistine COLIS

Martischang R et al. ICPIC 2019, late breaker (O45)

BACKGROUND



1



2



3

81105.0124/1	<p>ABS Klebsiella pneumoniae carbapenemase (KPC), par PCR</p> <p>PRES (^{na}New Delhi metallo-beta-lactamase (NDM), par PCR</p> <p>ABS (^{na}Oxacillinase 181 (OXA-181), par PCR</p> <p>ABS (^{na}Veronia integron-encoded metallo-beta-lactamase (VIM), par PCR</p> <p>ABS (^{na}Metallo-beta-lactamase IMP, par PCR</p> <p>ABS (^{na}SPM-type carbapenemase, par PCR</p> <p>ABS (^{na}GES-type carbapenemase, par PCR</p> <p>ABS (^{na}SME-type carbapenemase, par PCR</p> <p>ABS (^{na}GIM-type carbapenemase, par PCR</p> <p>ABS (^{na}SIM-type carbapenemase, par PCR</p>	(a)	+++ Escherichia coli +++ Klebsiella pneumoniae
181121.0612/1	<p>ABS Klebsiella pneumoniae carbapenemase (KPC), par PCR</p> <p>PRES (^{na}New Delhi metallo-beta-lactamase (NDM), par PCR</p> <p>ABS (^{na}Oxacillinase 181 (OXA-181), par PCR</p> <p>ABS (^{na}Veronia integron-encoded metallo-beta-lactamase (VIM), par PCR</p> <p>ABS (^{na}Metallo-beta-lactamase IMP, par PCR</p> <p>ABS (^{na}SPM-type carbapenemase, par PCR</p> <p>ABS (^{na}GES-type carbapenemase, par PCR</p> <p>ABS (^{na}SME-type carbapenemase, par PCR</p> <p>ABS (^{na}GIM-type carbapenemase, par PCR</p> <p>ABS (^{na}SIM-type carbapenemase, par PCR</p>	(c)	+++ Escherichia coli +++ Klebsiella pneumoniae
190503.0092/1	<p>ABS Klebsiella pneumoniae carbapenemase (KPC), par PCR</p> <p>PRES (^{na}New Delhi metallo-beta-lactamase (NDM), par PCR</p> <p>ABS (^{na}Oxacillinase 181 (OXA-181), par PCR</p> <p>ABS (^{na}Veronia integron-encoded metallo-beta-lactamase (VIM), par PCR</p> <p>ABS (^{na}Metallo-beta-lactamase IMP, par PCR</p> <p>ABS (^{na}SPM-type carbapenemase, par PCR</p> <p>ABS (^{na}GES-type carbapenemase, par PCR</p> <p>ABS (^{na}SME-type carbapenemase, par PCR</p> <p>ABS (^{na}GIM-type carbapenemase, par PCR</p> <p>ABS (^{na}SIM-type carbapenemase, par PCR</p>		1.0E4 Escherichia coli 1.0E2 Enterococcus
n de jet			



Probable transmission:

- 1st : Intermediate Care Unit (IMCU)
 - 2nd and 3^d : Private room

BACKGROUND

MET



ts' screening (July & Nov 18, May 19)

✓ 18 : Roomates screening (n=5) → negative

✓ 19 : Entire unit screening → negative

Environmental screening and disinfection in the private room (May 19)
Kitchen, bathroom, air conditioner : Negative (n=20)



- Illumina iSeq sequencing
- cgMLST and cgSNP analyses

Martischang R et al. ICPIC 2019, late breaker (O45)

METHODS & RESL

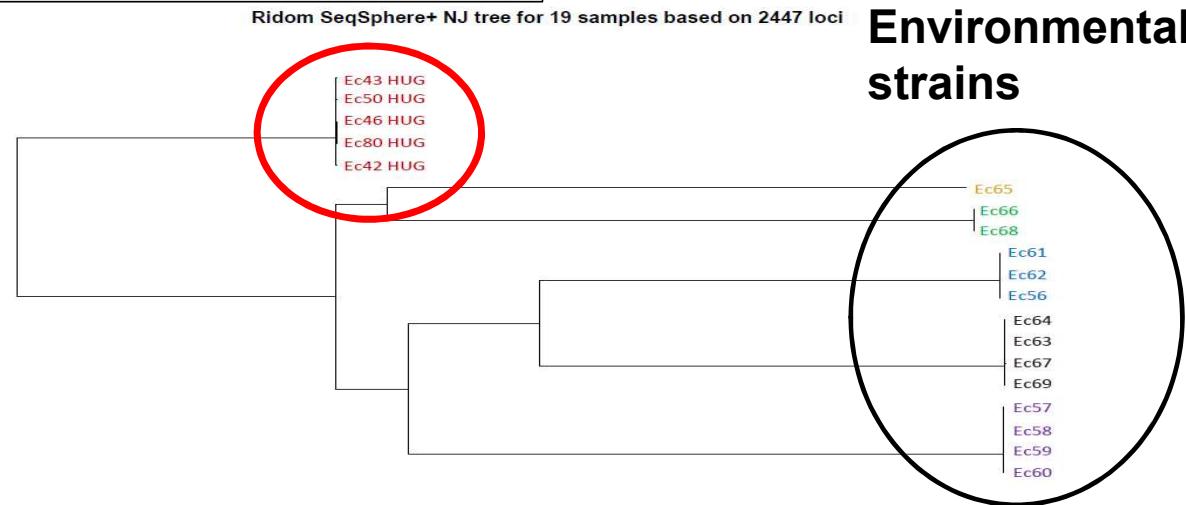
E. coli ST354 NDM-1 (<10SNPs)

- Mostly from animals
- Rarely associated with carbapenemases

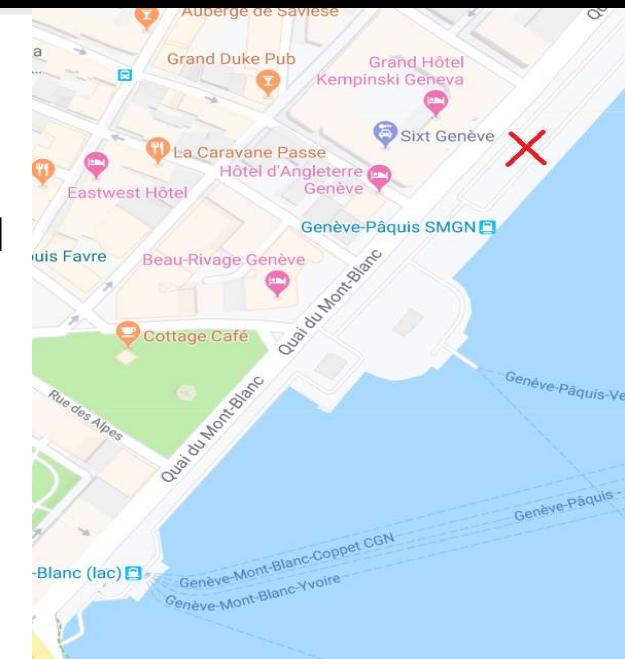
no missing values, % columns difference
Task Template: E. coli cg MLST v1.0, E. coli MLST Warwick v1.0
E. coli cgMLST Complex Type / Cluster-Alert distance: 10
Comparison Table Retrieval: Coli_19_souches [unstored]
Projects: Coli_19_souches (Escherichia coli)
Comparison Table created: May 16, 2019 2:27 PM (v5.1.0_(2018-06))
Distance based on columns from E. coli cg MLST (2447)

Sequence Types (ST):
● 354
● 940
● 361
● 38
● 405
○ No group assigned

Date 17/05/2019
Par Prof J. Schrenzel



- Reinforced attention on **standard precautions** in the concerned units.
- Implementation of a **computerized readmission alert system** (May 19) of all patients admitted in the unit from 11.18-05.19 (n=240) with a recommendation to **screen them** at re-admission (all are negative up to now).
- One of the patients **died of surgical complications** unrelated to *E. coli* NDM-1 carriage. There was no clinical infection related to *E. coli* NDM-1.



Courtesy: J Pot

RESULTS

1st healthcare-associated NDM-producing *E. coli* outbreak in Switzerland

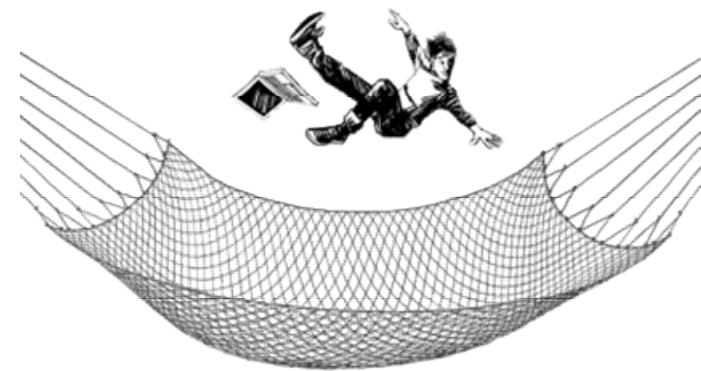
Prolonged cluster with hidden transmission detected by chance

Nosocomial transmission despite contact precautions for the index case

- Probable imperfect implementation of contact precautions
- Private floors are not exempt from MDROs nosocomial acquisition and may be at high risk of importation

Highlights the importance of :

- **Added value of NGS to guide investigation of an outbreak**
- **Environmental hygiene**
- **Admission screening for patients at-risk**
- **Weekly screening in ICU (shared benefits with other units/wards)**



General internal medicine ward, Geneva

Sporadic, imported CPE
(OXA-48 cluster in 2011)



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KPC outbreak in Geneva, 2015

- Mr CV, Italian origin, known KPC carrier since 2012
- Admitted in January 2015 for severe KPC urosepsis
- Control measures were applied (private room)



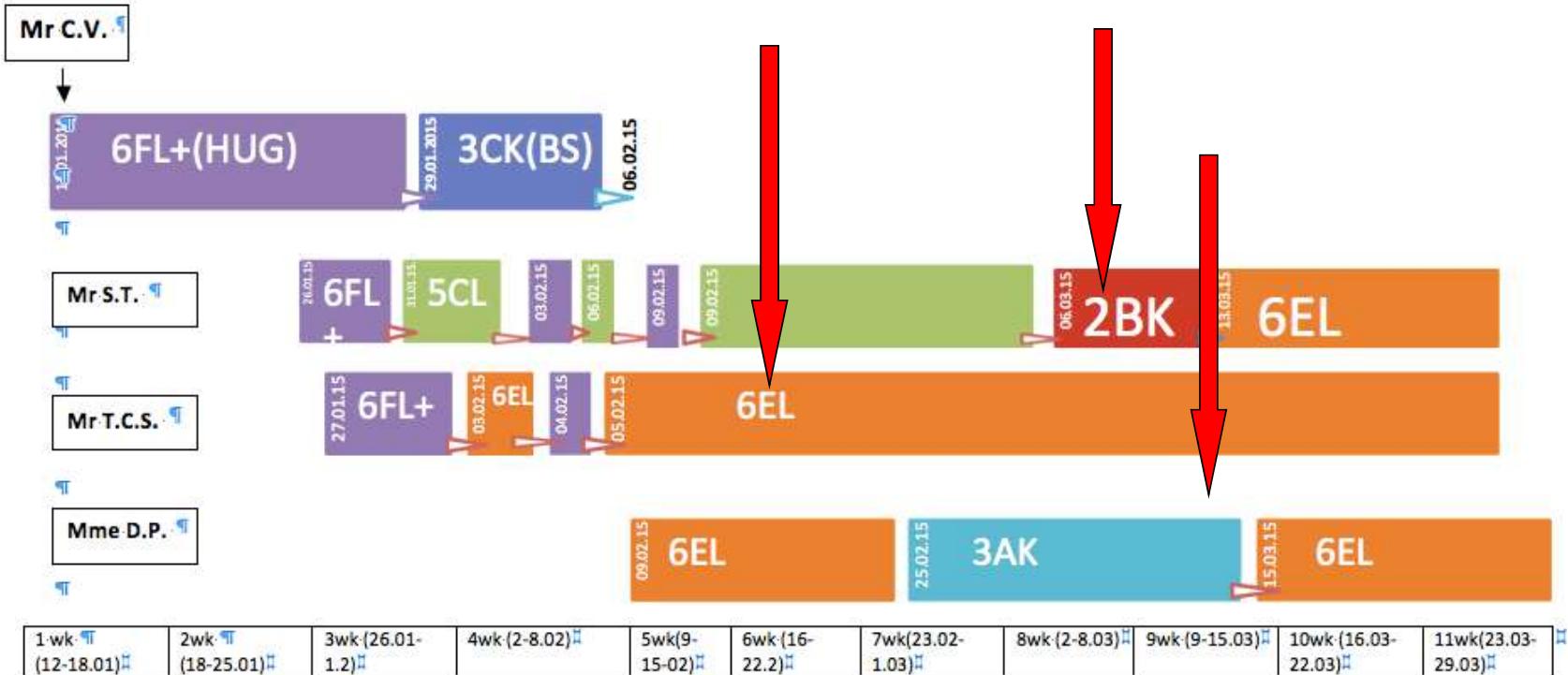
Courtesy: F. Olearo, D. Pires



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Spread of KPC



Courtesy: F. Olearo, D. Pires



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Voir sous MS-Excel	06/03/2015 10:25:00	04/03/2015 11:45:00	04/03/2015 11:45:00	27/02/2015 19:00:00	27/02/2015 19:00:00	20/02/2015 17:00:00	19/02/2015 10:55:00	16/01/2015 18:15:00	16/01/2015 18:10:00	16/01/2015 18:10:00	12/01/2015 12:15:00	27/12/2014 21:15:00	04/09/2014 12:00:00	21/07/2014 15:45:00	14/04/2014 06:50:00	
Klebsiella pneumoniae	Klebsiella pneumoniae	Klebsiella pneumoniae	Proteus mirabilis	Klebsiella pneumoniae	Proteus mirabilis	Klebsiella pneumoniae	Klebsiella pneumoniae	Bacillus cereus	Pseudomonas aeruginosa	Proteus mirabilis	S. aureus méticilline-sensible (MSSA)	S. aureus méticilline-sensible (MSSA)	Escherichia coli	Escherichia coli	Pseudomonas aeruginosa	Escherichia coli
sgv ponction	frot. plaq prof.	frot. plaq prof.	frot.	frot.	liqu.	liqu.	frot. plaq prof.	frot. plaq prof.	frot. plaq prof.	frot. plaq prof.	tissu mou	urine	urine	frot. escarre	urine	
Penicilline G							(32) R			R	R					
Flucloxacilline										S	S					
Amoxicilline	R	R	R	R	R	R			R			R	R		R	
Co-amoxiclav	R	R	S	R	S	R			S	S	S	S	S		S	
Piperacilline	R	R	S	R	I	R	R		R	S					S	
Piperac.+tazob	R	R	S	R	S	R	R		S	S		S	S	S	S	
Cefalotine																
Cefoxitine	R	R	S	R	S	R	R		S			S	S		S	
Cefuroxime	R	R	S	R	S	R	R		S			S	S		S	
Cefuroxime axetil (oral)																
Ceftazidime	R	R	S	R	S	R	R		S	S		S	S	S	S	
Ceftriaxone	R	R	S	R	S	R	R	(32) I	S			S	S		S	
Cefepime	R	R	S	R	S	R	R		S	S		S	S	S	S	
Imipenem	(>32) R	R	S	(32) R	(0.500) S	R	(32) R	(32) R	I	S		S	S	S	S	
Meropeneme	(>32) R	R	S	(32) R	(0.032) S	R	(>32) R	(>32) R	I	S		S	S	S	S	
Ertapénem	(>32) R	R	S	(32) R	(0.160) S	R	(>32) R		S			S	S	S	S	
Aztreonam	R	R	S	R	S	R	R		S	S						
Amikacine	R	R	S	R	S	R	I		S	S		S	S	S	S	
Gentamicine	S	S	R	S	R	S	S		S	S	S	S	S	S	S	
Tobramycine									S						S	
Norfloxacine	R	R	S	R	S	R	R		R			R	R	R	R	
Ciprofloxacine	R	R	S	R	S	R	R		S	R	S	S	R	R	S	
Levofloxacine									S						S	
Clindamycine								(1) I				S	S			
Erythromycine												S	S			
Acide fusidique												S	S			
Co-trimoxazole	R	R	R	R	R	R	R		S	S	S	R	R		R	
Fosfomycine	R	R	S	S	S	R	R		S		S	S	S		S	
Rifampicine																
Tetracycline																
Tigécycline	S	S	R	S	R	S	S		R	S	S					
Vancomycine									(3) S			S	S			
Teicoplanine												S	S			
Linézolid												S	S			
Mupirocine												I				
Furanes	R	R	R	R	R	R	R		R			S	S		S	
Colistine	(12) R	(4) R		(16) R			(0.750) S								S	
Carbapénémase-OXA48								A								
Carbapénémase-KPC								P								

Total: 3 cases of KPC cross-transmission

(2 clinical infections + 1 asymptomatic colonization)



Need of a bundled intervention

- Contact tracing with widespread screening
- Cohorting / strict contact precautions
- Electronic re-admission alert system
- Information (HCWs, patients, families)

Brochure

- A booklet with information about KPC addressed to the patients at risk and their families

Klebsielle KPC : une bactérie multi-résistante

De qu'il faut savoir sur cette entérobactéries : comment la dépister et comment éviter sa transmission ?



Qu'est-ce qu'une Klebsielle KPC?

Nous sommes tous naturellement porteurs de bactéries sur notre peau, nos muqueuses et dans notre tube digestif. La plupart répondent aux antibiotiques, mais certaines deviennent résistantes. C'est le cas de la bactérie Klebsielle KPC.

Quelle est la particularité d'une Klebsielle KPC? Cette bactérie fait partie de la flore digestive et appartient à la classe des entérobactéries. Avec l'usage des antibiotiques, elle peut devenir hautement résistante et produire des enzymes (Klebsiella Pneumoniae productrice de Carbapénémases = KPC) qui inhibent l'effet de la quasi-totalité des antibiotiques habituels, dits à large spectre.

Comment se transmet cette bactérie ? La transmission se fait par voie oro-faciale lors de contacts directs ou indirects par l'intermédiaire des mains, d'aliments, d'eau, de surfaces contaminées.

Comment sait-on que l'on est porteur ? Il faut réaliser un dépistage car une personne peut être porteur sans avoir de symptômes. Cette bactérie peut disparaître spontanément ou rester plusieurs semaines ou mois dans le tube digestif sans provoquer une infection. Parfois une infection digestive ou urinaire survient et son traitement avec des antibiotiques est alors difficile.

Comment se fait un dépistage ? Le dépistage consiste en un frottis rectal ou un prélevement de selles qui sont analysés dans un laboratoire de bactériologie. Le résultat est disponible dans les 48 à 72 heures.

Quel est l'intérêt de faire un dépistage ? Si le dépistage est positif, votre médecin sait que vous êtes porteur de la Klebsielle KPC. Si vous contractez une infection, il prend contact avec un service spécialisé pour vous prescrire un traitement antibiotique adapté.

Que faire si vous êtes porteur d'une Klebsielle KPC?

Voici les précautions à prendre chez vous pour éviter de transmettre cette bactérie à vos proches.

- Utilisez votre savon habituel et lavez-vous les mains chaque fois que cela est nécessaire :
 - après être allé aux toilettes
 - avant de préparer des repas et de passer à table
 - après avoir manipulé une poche à urine, du linge souillé ou un pansement.
- Faites votre toilette tous les jours en utilisant votre savon habituel, mais ne partagez pas votre linge de toilette.
- Lavez votre linge, à une température de 40°C minimum, en utilisant votre produit à lessive habituel.
- Utilisez vos produits de nettoyage habituels pour le ménage.
- Nettoyez vos sanitaires et votre salle de bains avec votre détergent habituel et rincez bien.
- Entretenez normalement votre vaisselle et vos ustensiles de cuisine.
- Eliminez vos pansements et vos protections souillées avec les ordures ménagères en fermentant hermétiquement les sacs poubelles; n'oubliez pas de vous laver les mains après.

Y a-t-il des risques pour vos proches ? Si vous suivez les précautions ci-dessus, le risque de transmission à votre famille ou vos proches est extrêmement faible. Si vous avez des personnes fragiles dans votre entourage (infirmier, femme enceinte, malade chronique...), demandez conseil à votre médecin.

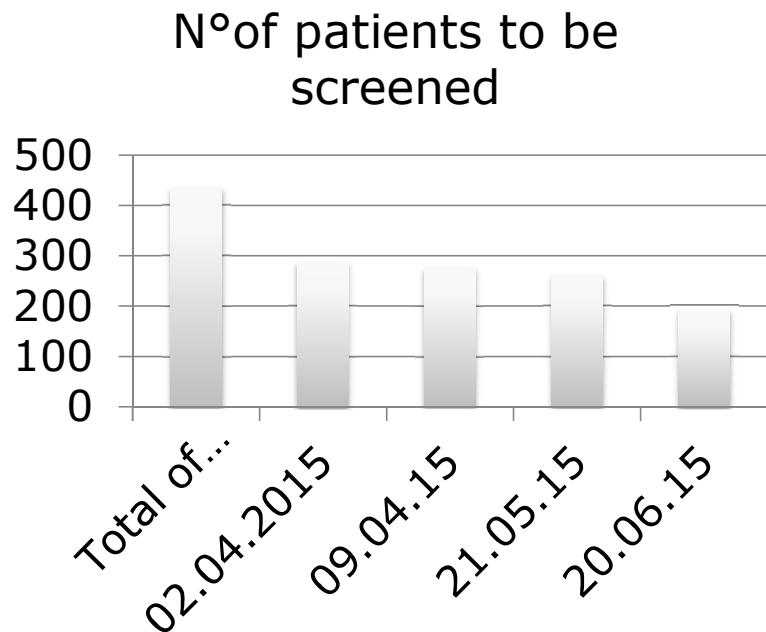
Courtesy: F. Olearo, D. Pires



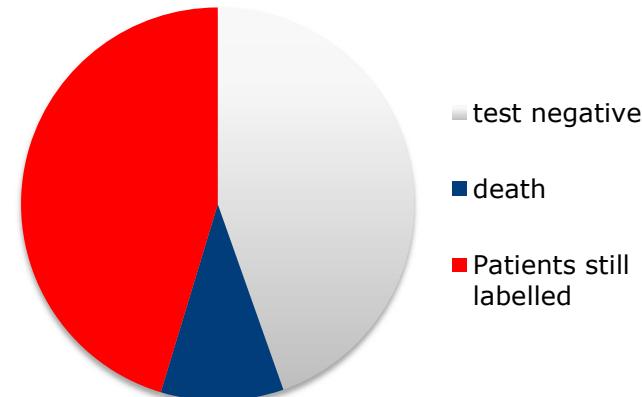
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Results of extensive screening



Screening test results



Not one single additional KPC case detected!

Courtesy: F. Olearo, D. Pires

Lessons learned (CPE outbreak)

- Contact precautions failed for previously identified KPC index case
- Late outbreak detection
- Screening program: logistical challenge

- WGS:
 - In slight contradiction to strong epidemiological evidence
 - Interesting post-hoc exercise

Serratia outbreak

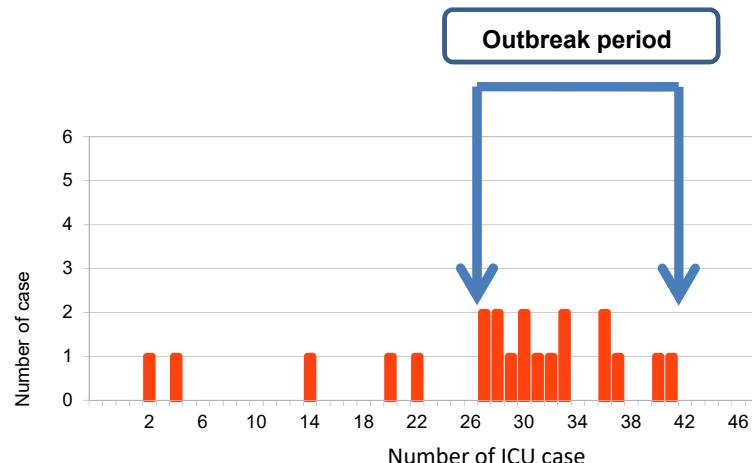
Geneva ICU



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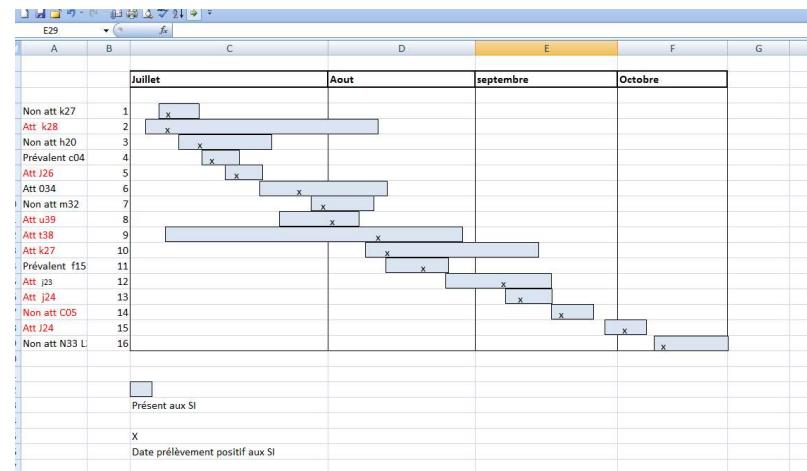
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Routine Serratia Surveillance => Epidemic curve in the ICU



HH compliance: 66 % (1st semestre) to 52% (3^d Trimestre)

Epidemiologic outbreak investigation => nosocomial transmission?



Genomic investigation

-Selection and sequencing of multiple *S.marcescens* isolates (incl. outbreak strains) stored in the microbiological laboratory



Aligner le texte · Convertir en graphique SmartArt · Organiser Styles rapides · Contour · Recadrer · Remplacer · Sélectionner · Modification

Paragraphe

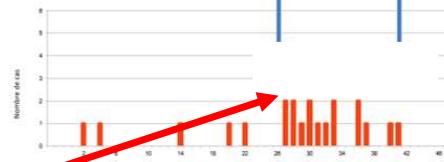
Dessin

Effets ·

Recadrer · Sélectionner · Modification

Courbe épidémique

Période épidémique



Bilan des interventions (5)

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Paragraphe

Dessin

Effets ·

Recadrer · Sélectionner · Modification

Volet Animation

Lire tout

Coller

Reproduire la mise en forme

Presse-papiers

Sequençage (RISQOM)

référence. Au total

(Minimum Spanning

Nos résultats mon

Cluster 2
30.12.201

Cluster 2
(4.7.2017,

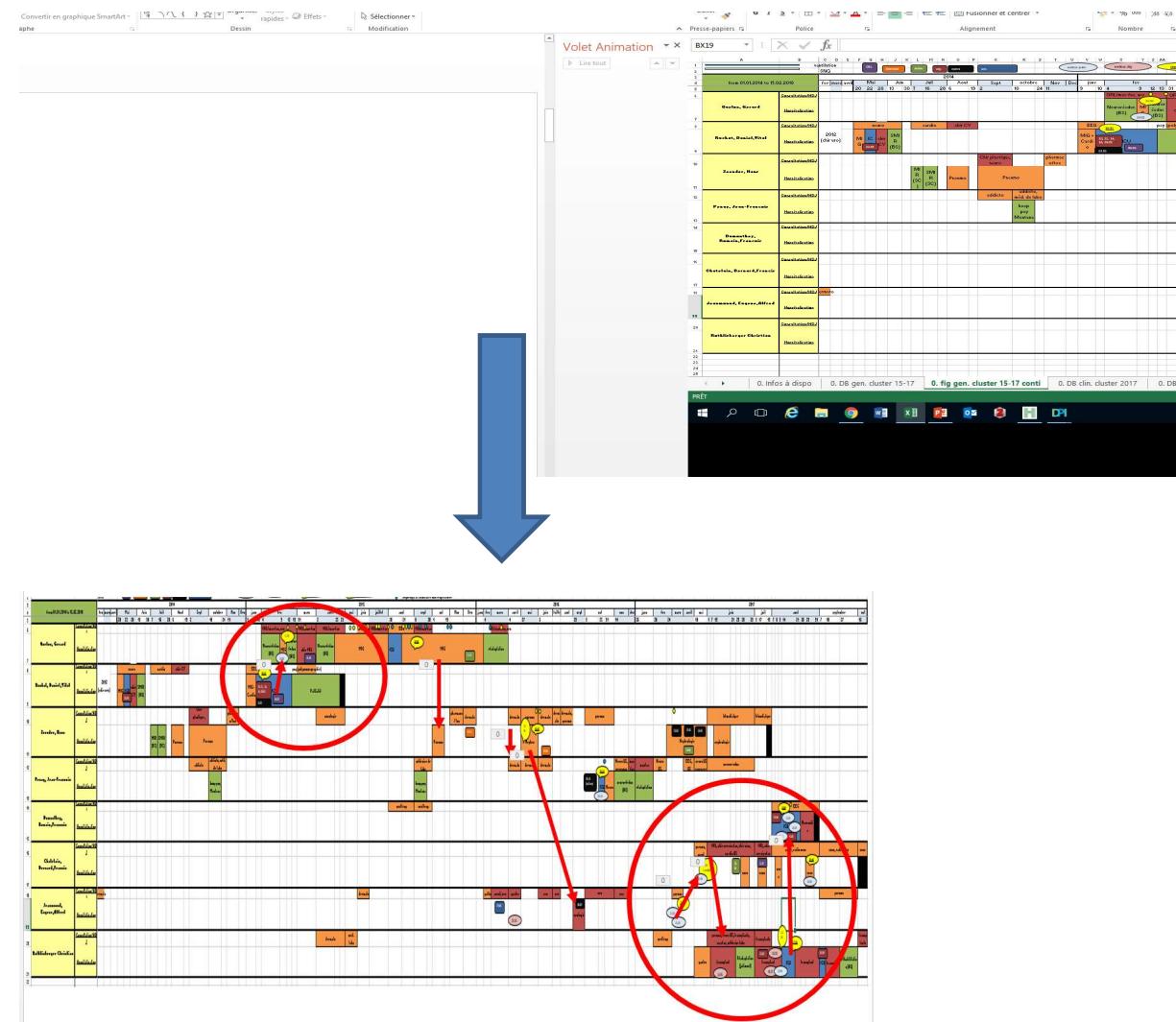
Cluster 1

Epidemiologic investigation

- Data collection to retrieve epidemiological links based on genomic data (small monoclonal cluster from 2015 to 2017)
- Epidemiological investigation based on geospatial, microbiological and medical information (respiratory procedures, respiratory therapy, surgery etc...)

BUT

- Selection & detection bias (missing cases)
- Misclassification bias
- Information bias (retrospective study)



Conclusions



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Required improvements to help hospital epidemiologists

WGS can reveal detailed spatial and temporal dynamics of nosocomial transmission events and MDRO evolution, but we need:

- Quality standards, proficiency testing for routine use
 - Standard-operating procedures for sampling, data extraction...
 - Thresholds to determine clustering and transmission events
 - Shorter TAT
 - Reduced costs
 - Streamlined data analyses
-
- **Most importantly: demonstrate impact of WGS on preventive measures and clinical decision making** (compared to less expensive tools)

Challenges of a paradigm shift: still not prime time !

- **Surveillance strategy**

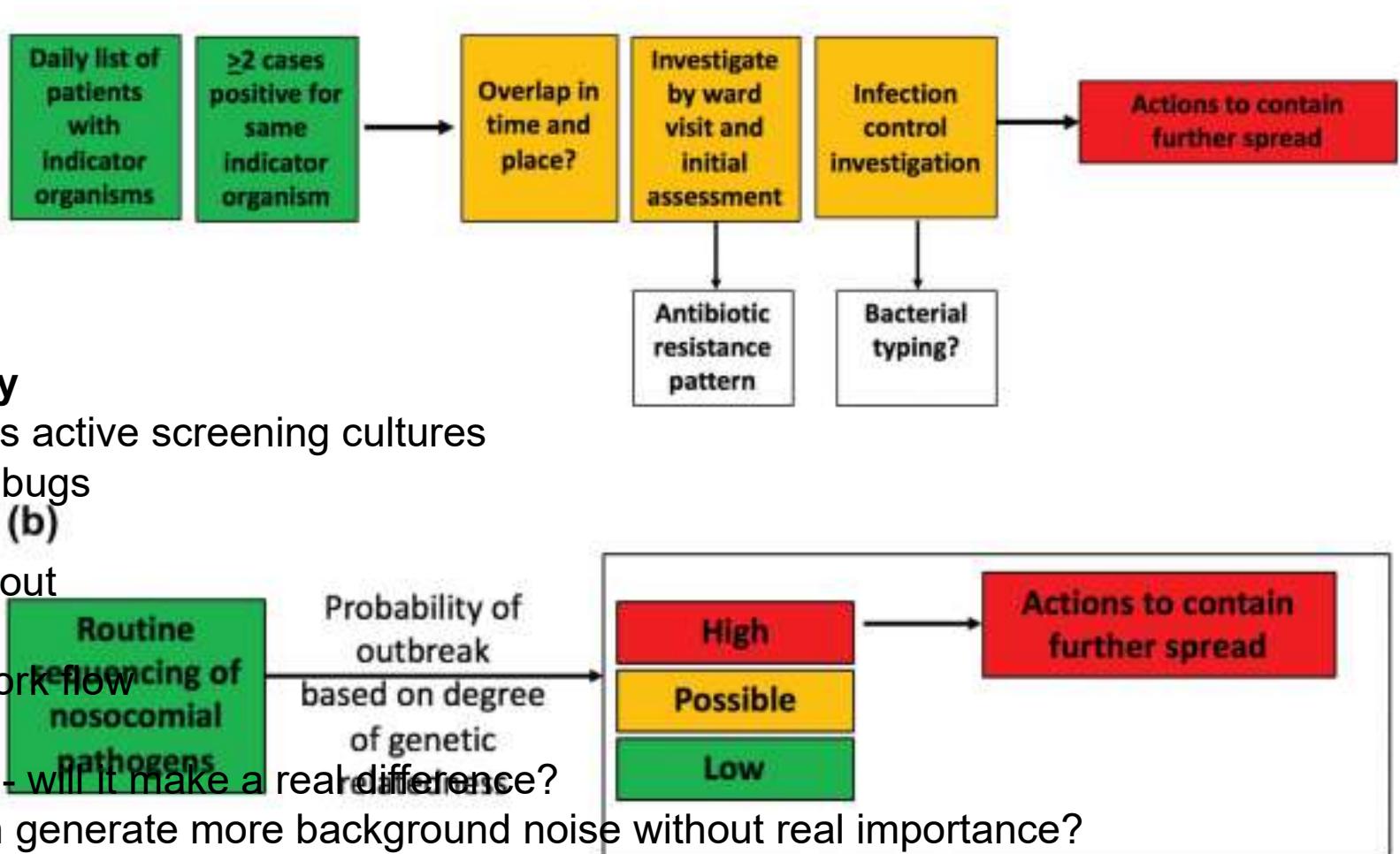
- Clinical cultures vs active screening cultures
- All bugs vs some bugs

- **Implementation (b)**

- In-house vs farm-out
- Cost
- Integration into work flow

- **Challenges**

- Turn around time - will it make a real difference?
- Will this approach generate more background noise without real importance?





Merci bcp pour
votre attention!

Reserve slides



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Challenges of a paradigm shift

- **Surveillance strategy**

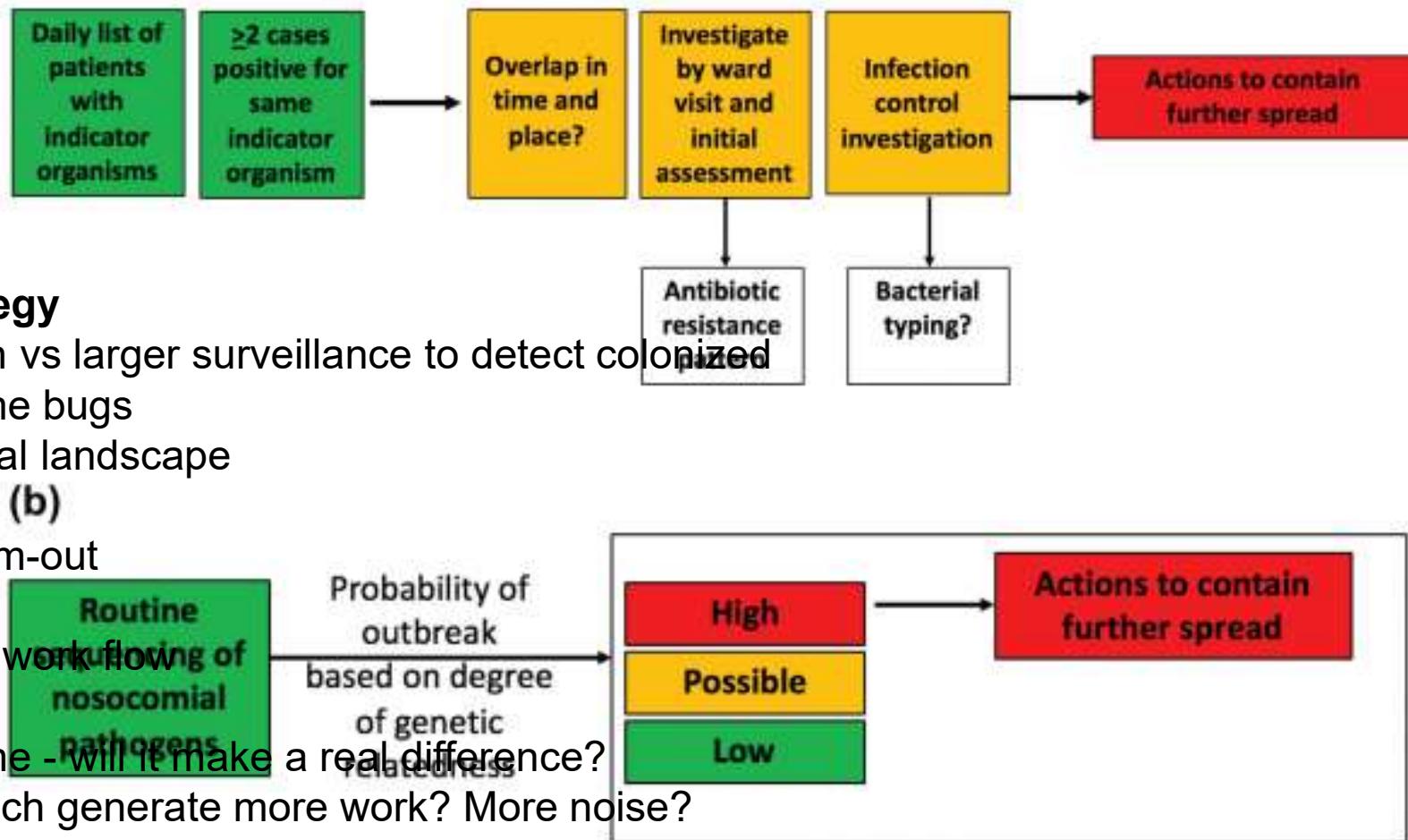
- Clinically-driven vs larger surveillance to detect colonization
- All bugs vs some bugs
- Hospital-regional landscape

- **Implementation (b)**

- In-house vs farm-out
- Cost
- Integrating into workflow

- **Challenges**

- Turn around time - will it make a real difference?
- Will this approach generate more work? More noise?



Do we have an outbreak?

Epidemiological question & hypothesis
Adequate screening strategy
Correct sampling & culturing practices

Choice of typing method & platform
Be aware of multiple biases during the process
Quality control issues

Analytical approach & interpretation

Allelic profile (ST)

SNPs profile

Resistance genes

Phylogenetic trees

Epidemiologic investigation

Threshold interpretation

Future improvements to help clinicians & hospital epidemiologists

- Standardization of:
 - Quality standards, (in-silico) proficiency testing
 - Standard-operating procedures for sampling, data extraction...
 - Global consensus on a set of cluster complexes specific genomes used as to call SNPs within ST- or CC groups.
 - Nomenclature (for databases and reference genomes)
- Research agenda:
 - Thresholds to determine clusters complexes according to different pathogens
 - Comparison hqSNP, cg-MLST and wg-MLST
 - Whole genome sequencing approaches vs robust epidemiologic data
 - Automatic curation of databases (for cgMLST)
- Agreements on data sharing practices (larger outbreaks)

